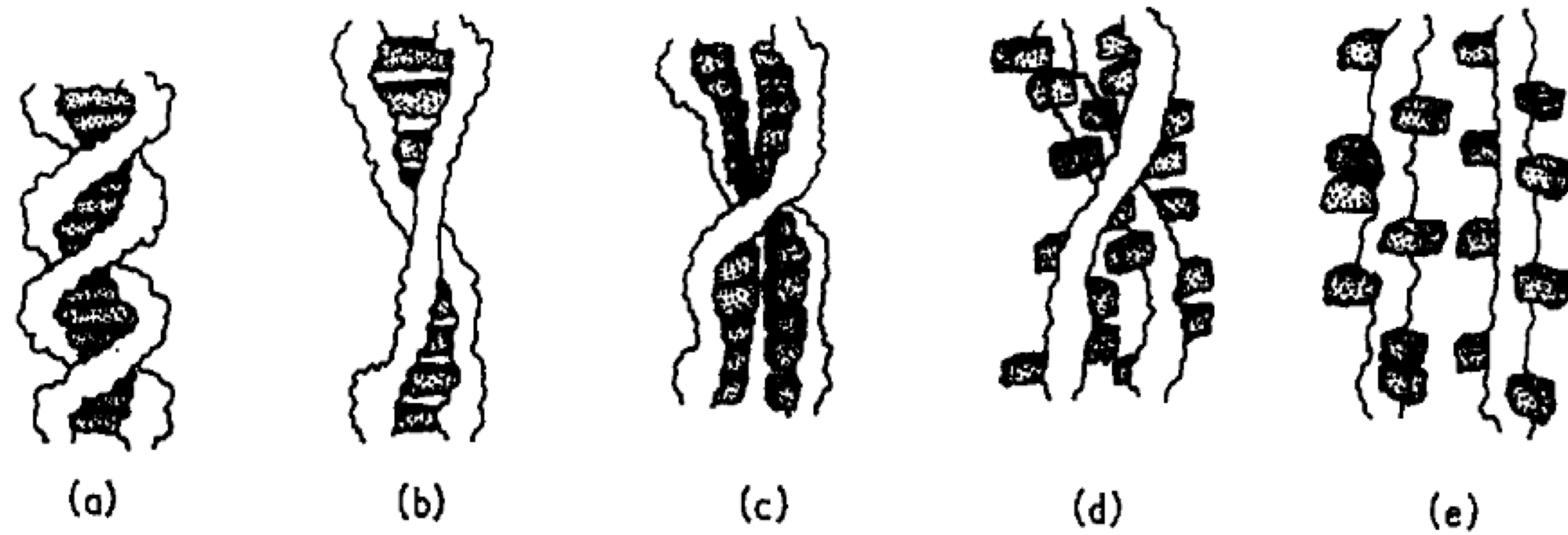


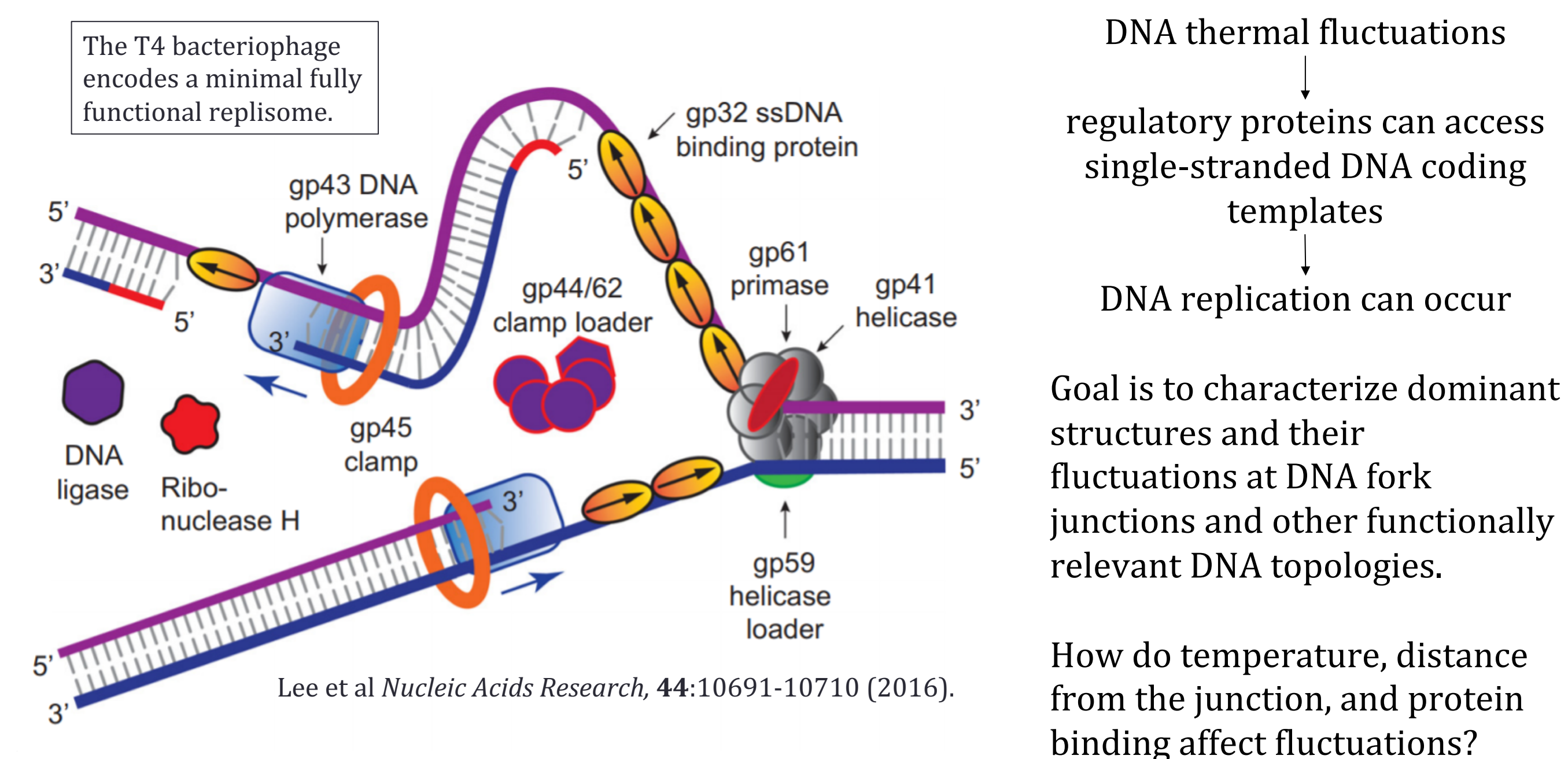
DNA "BREATHING"



McConnell, von Hippel *J. Mol. Biol.*, **50**: 298 (1970).

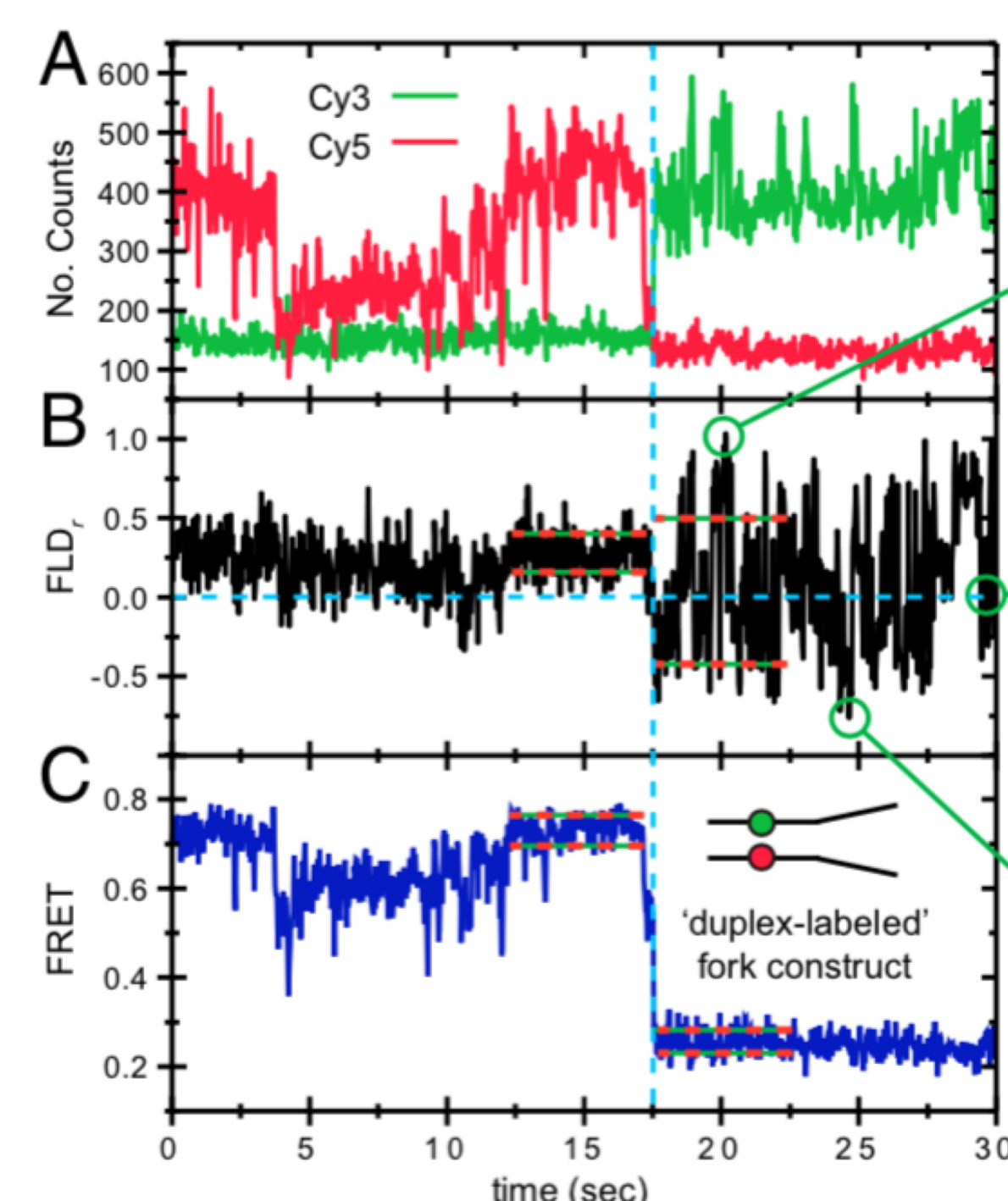
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Lee et al *Nucleic Acids Research*, **44**:10691-10710 (2016).

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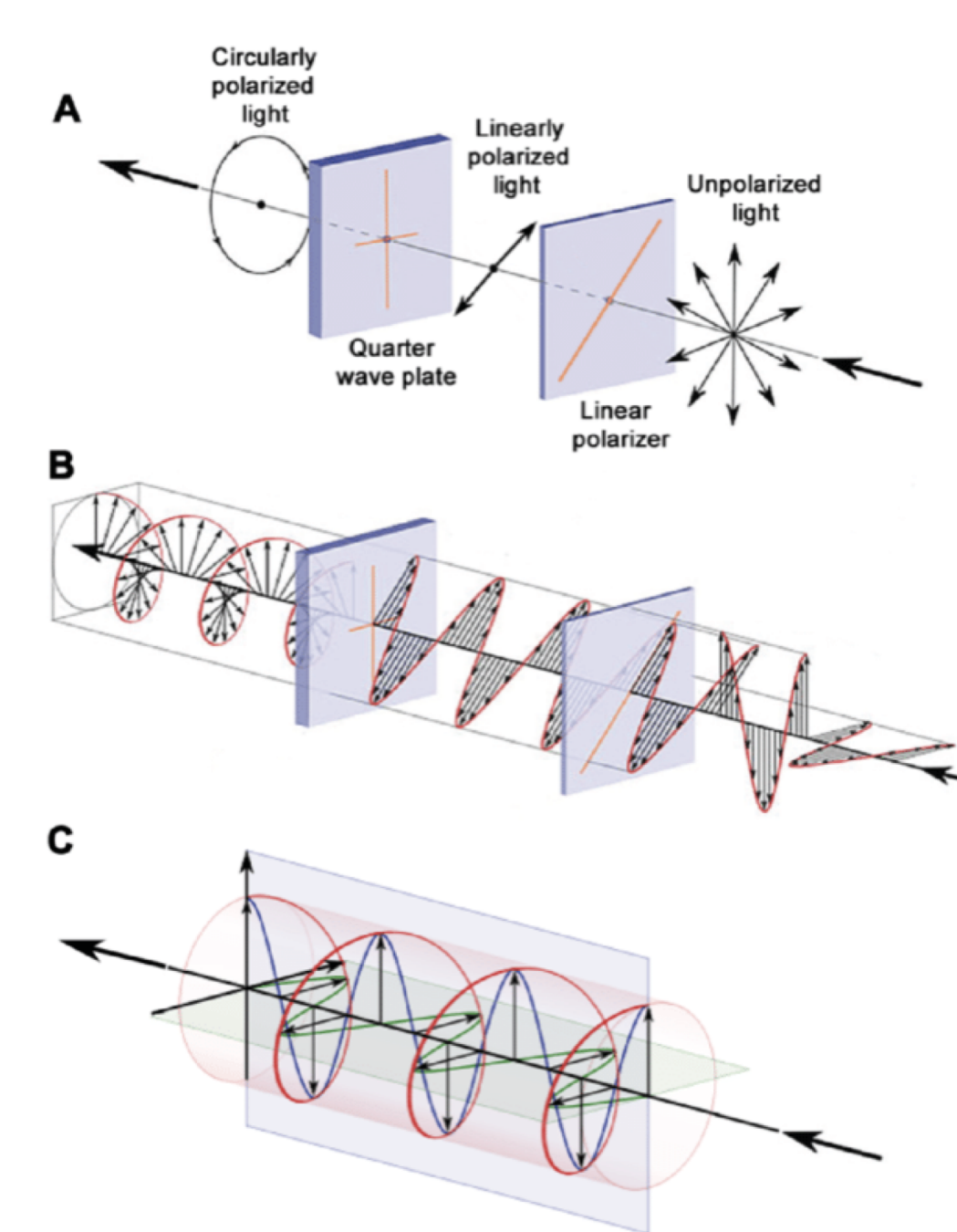
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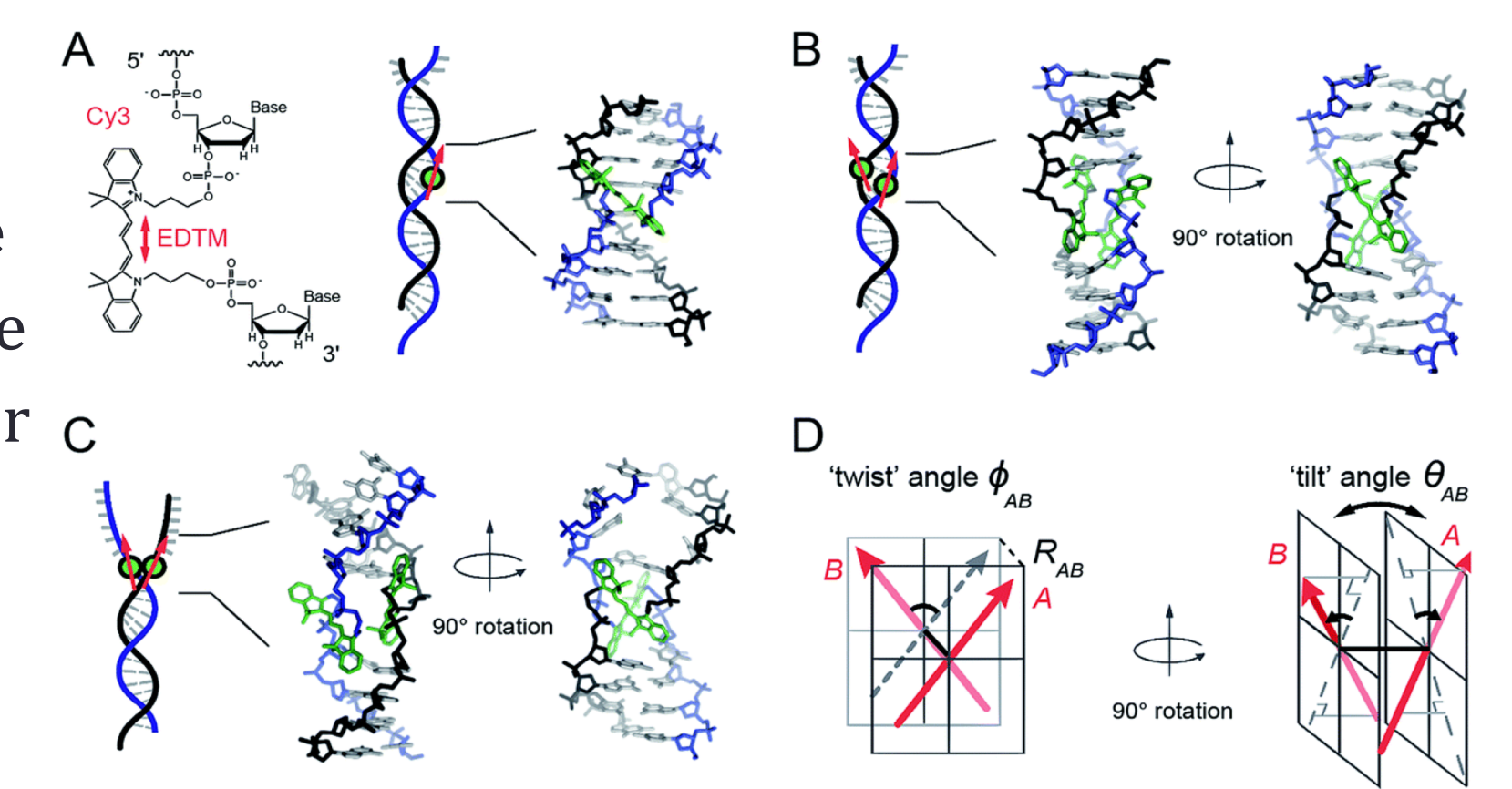
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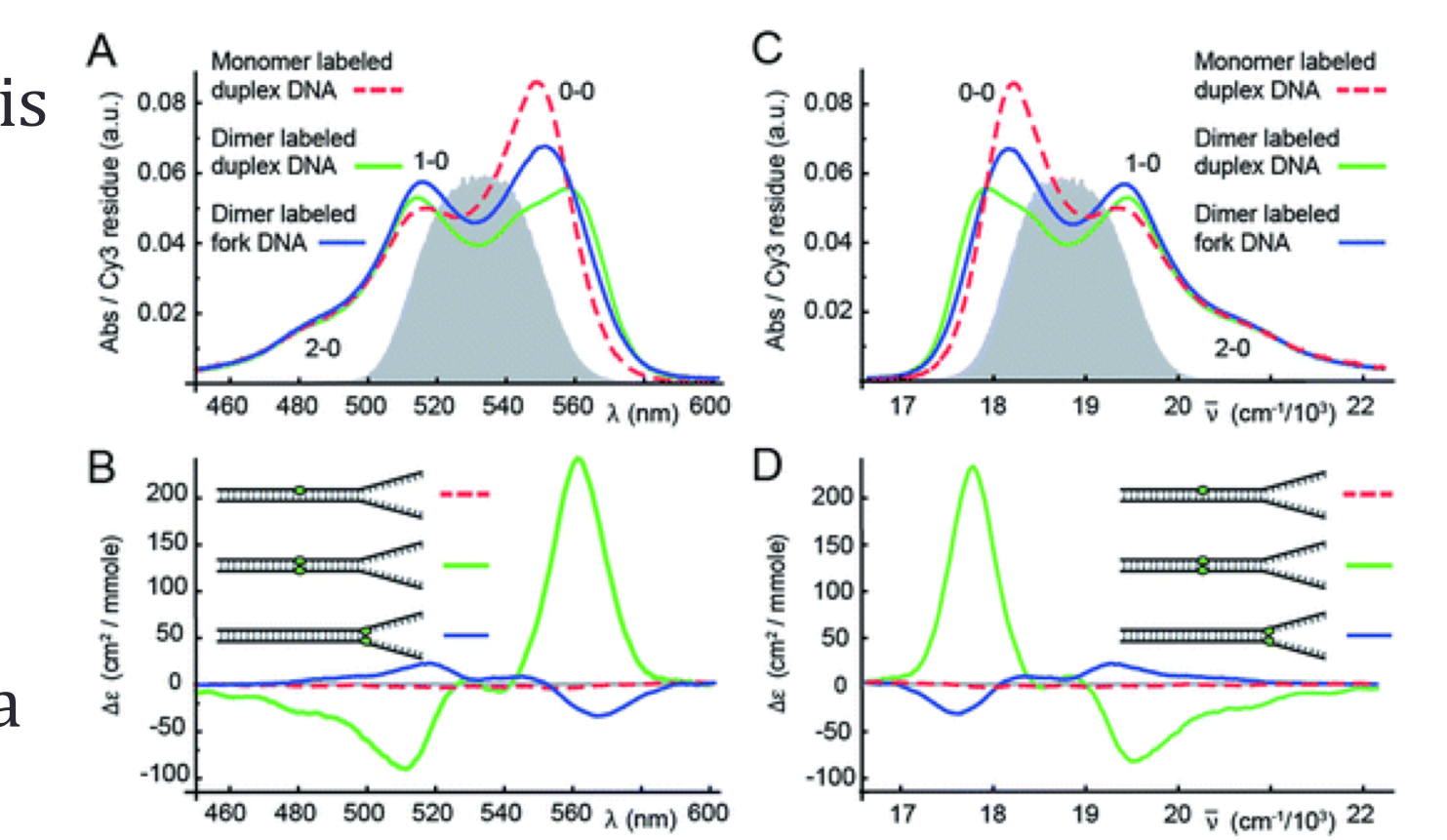


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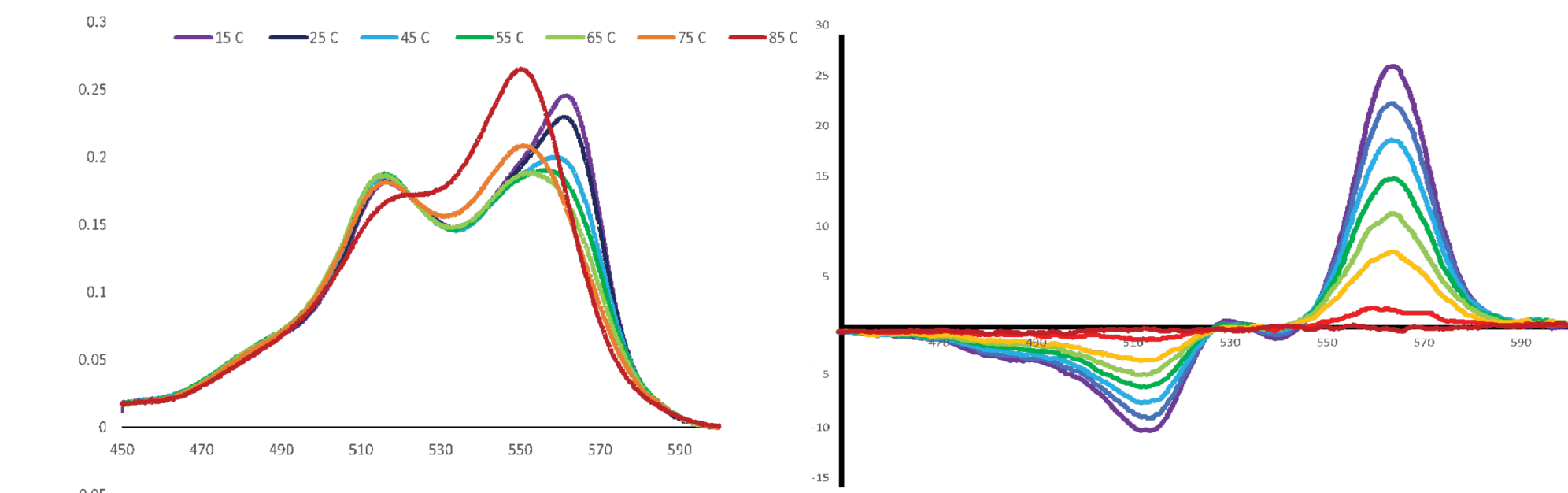
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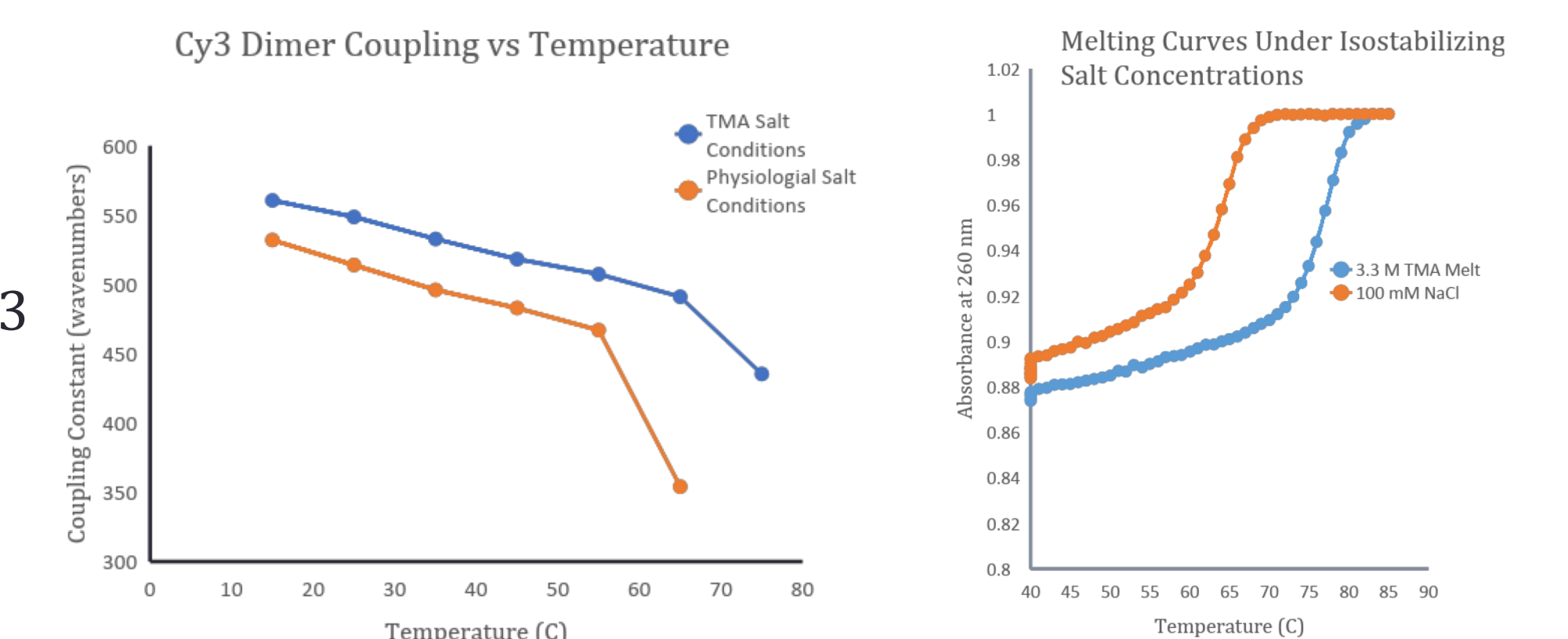
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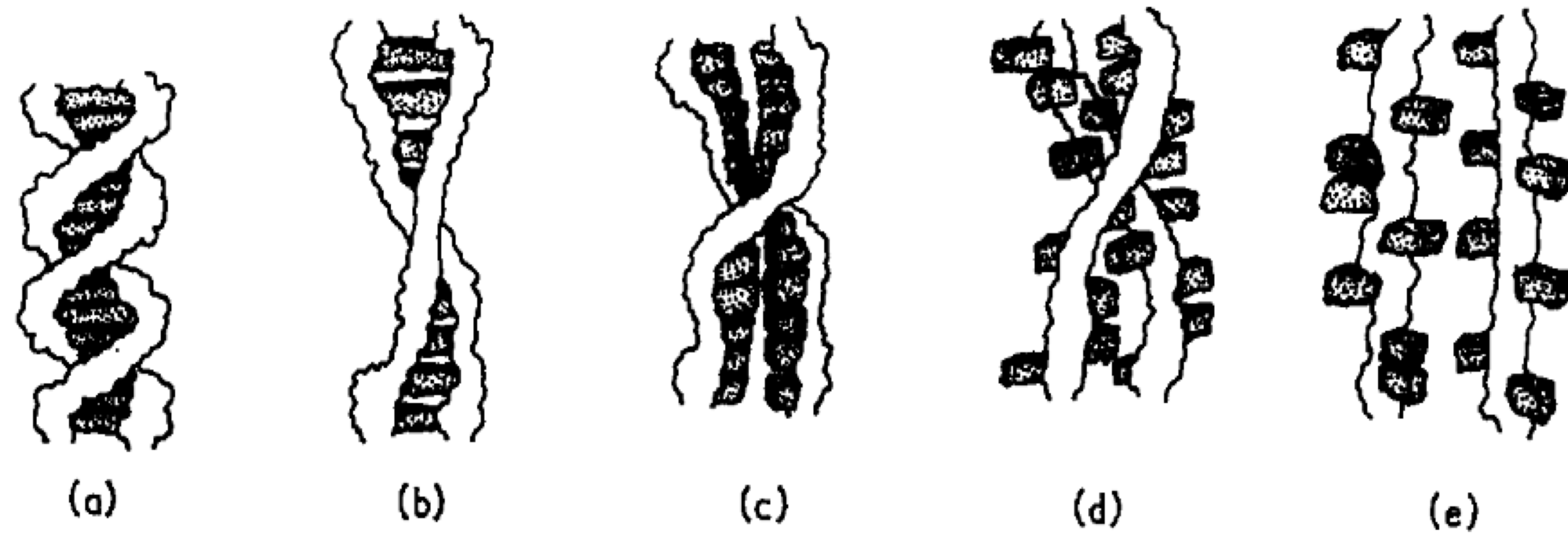
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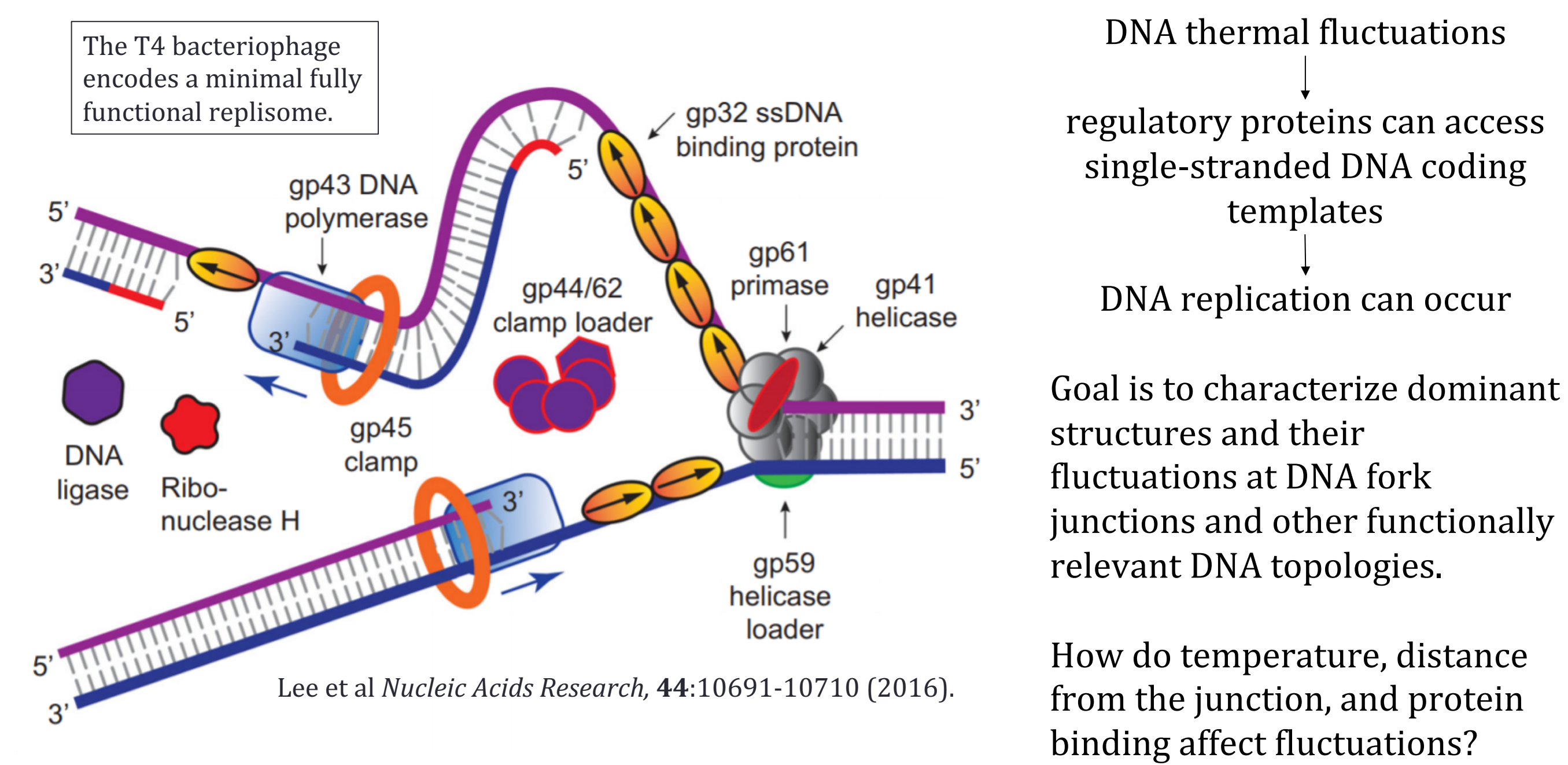
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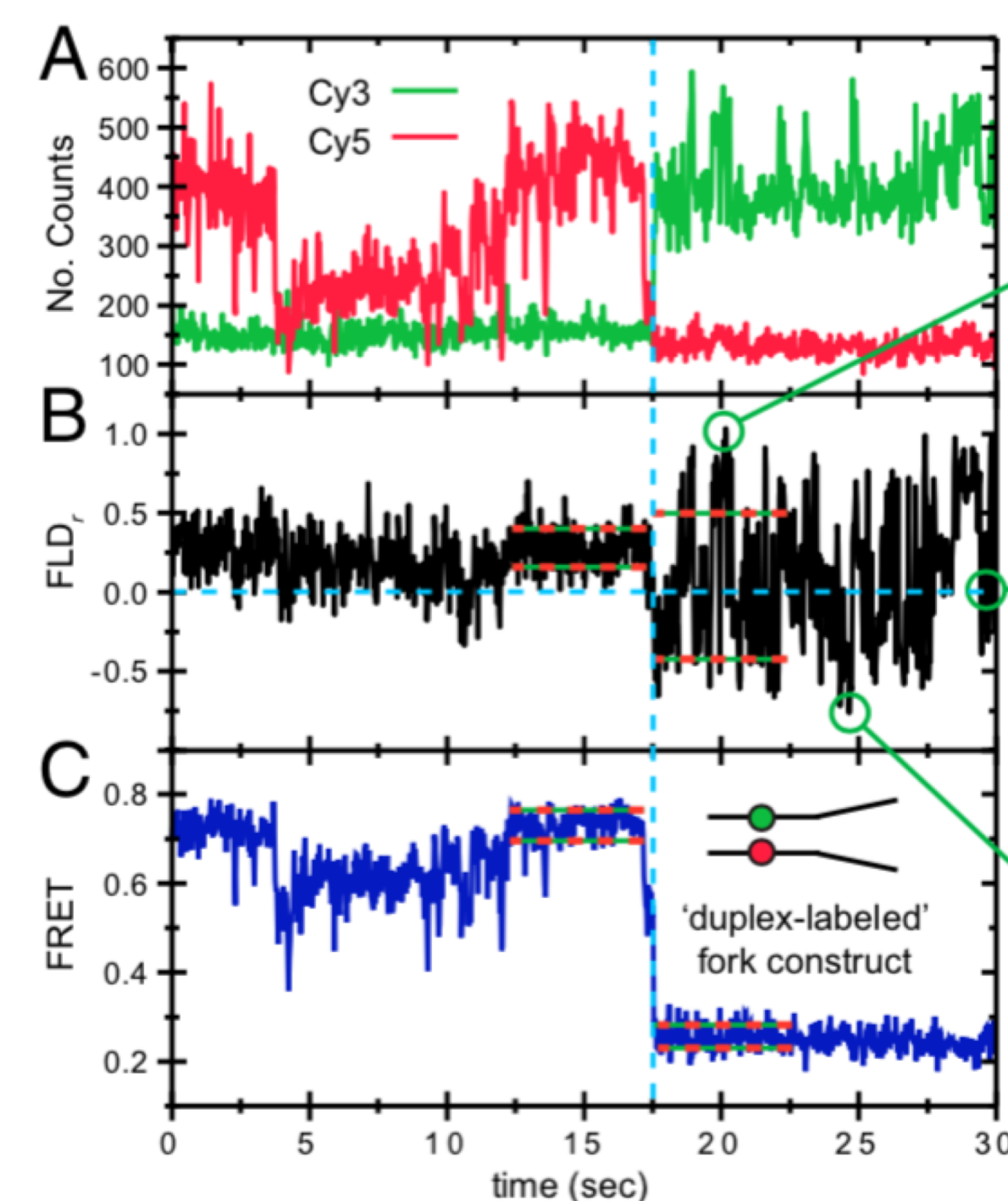
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How do temperature, distance from the junction, and protein binding affect fluctuations?

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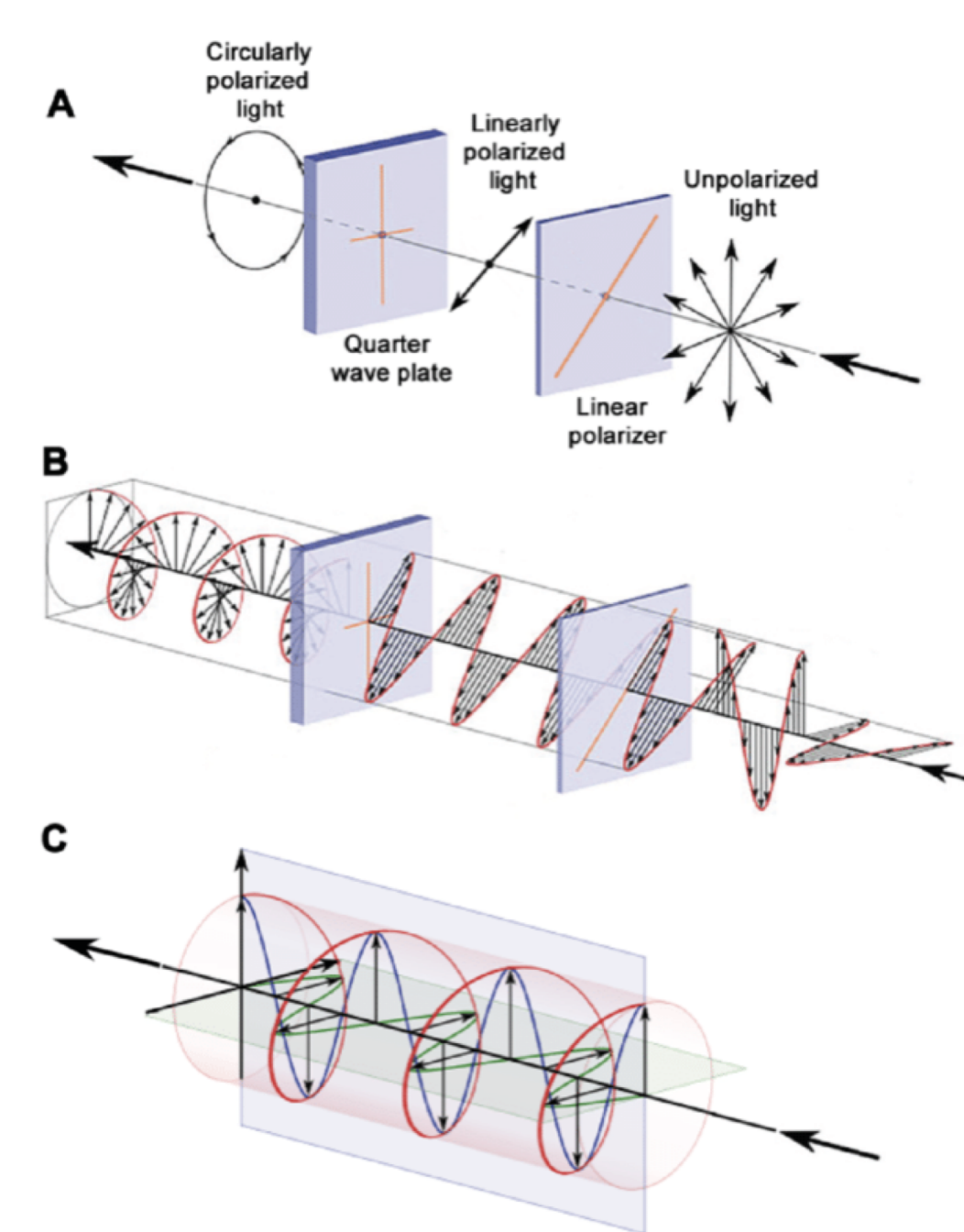
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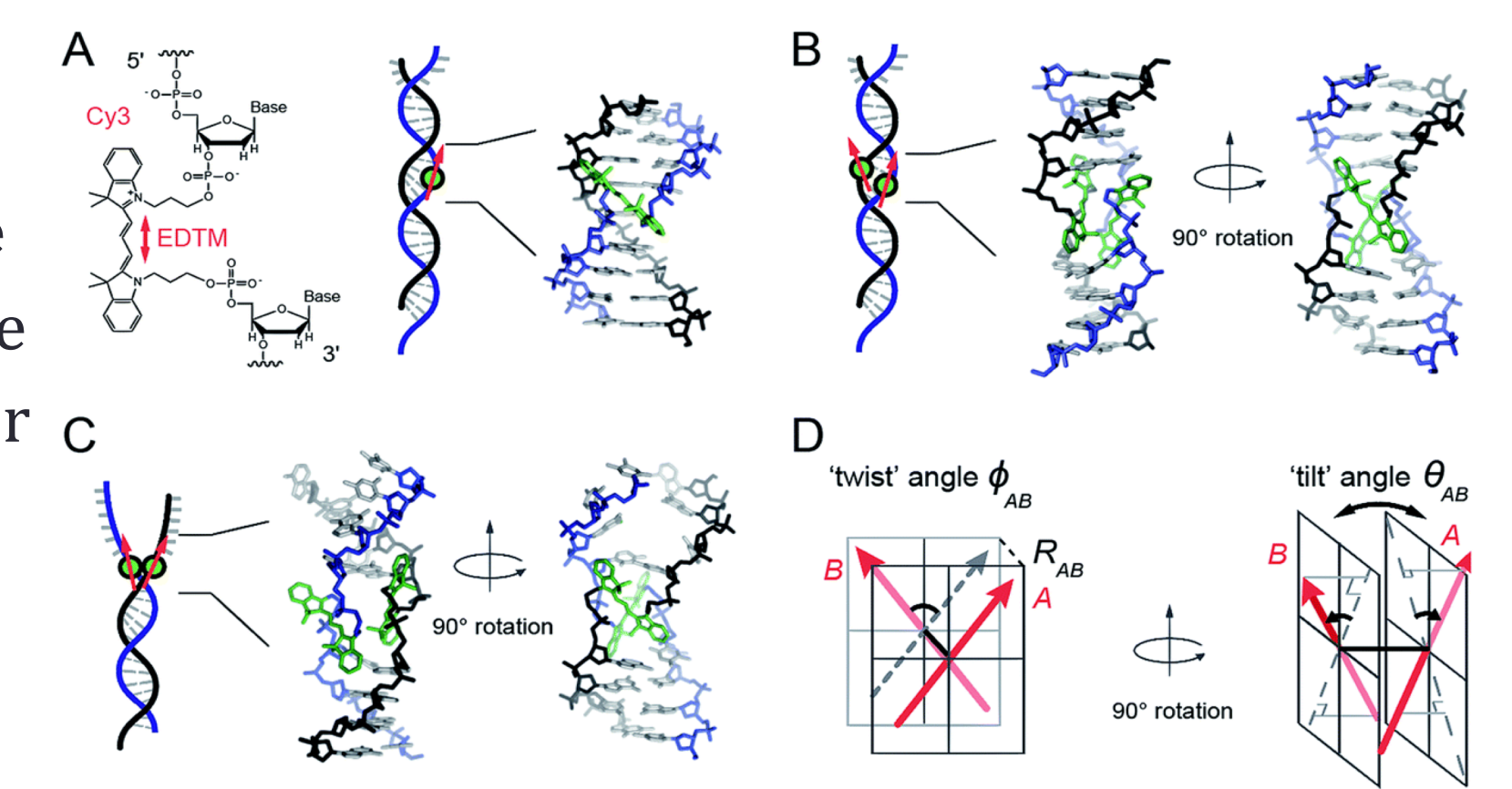


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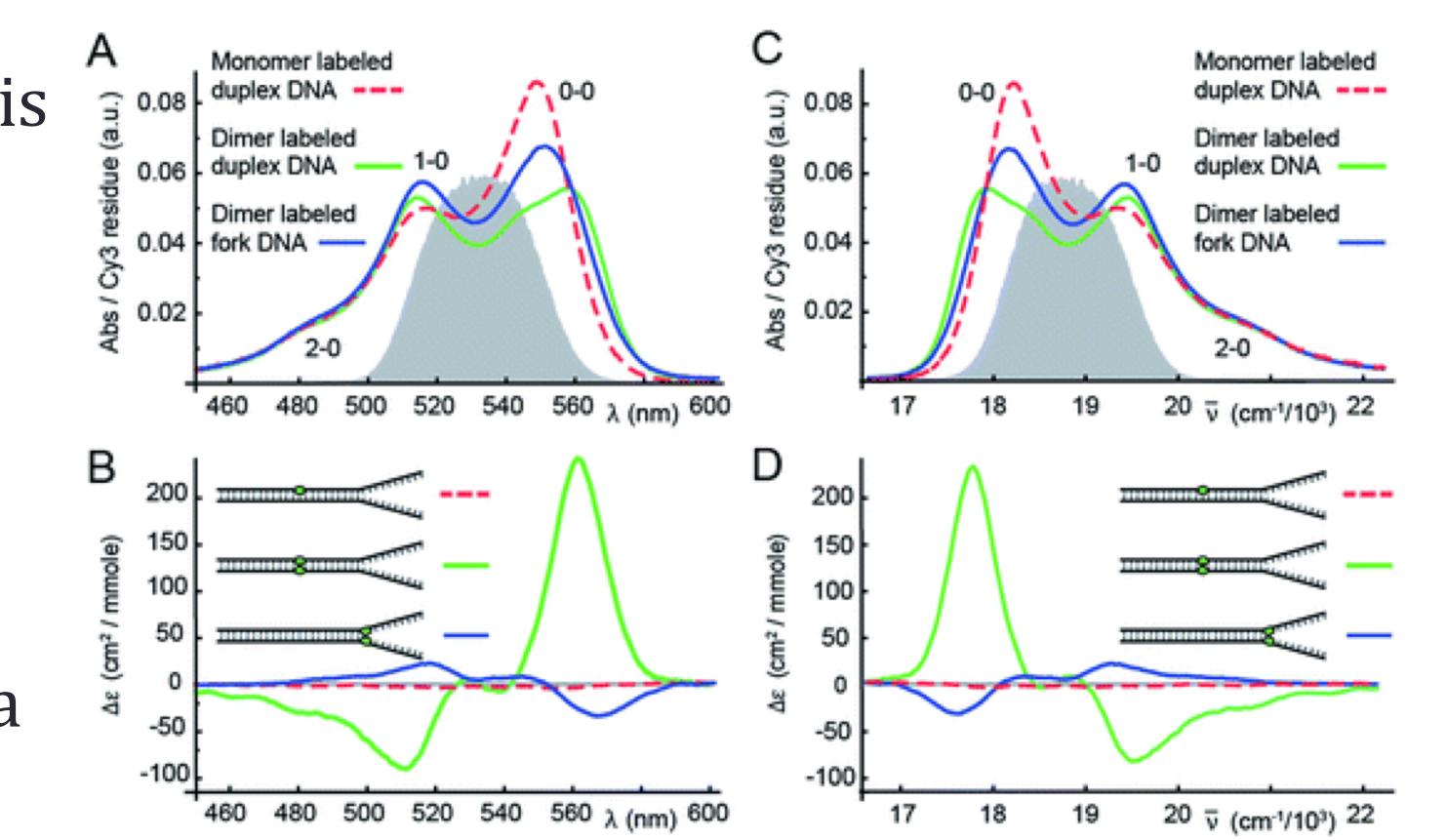
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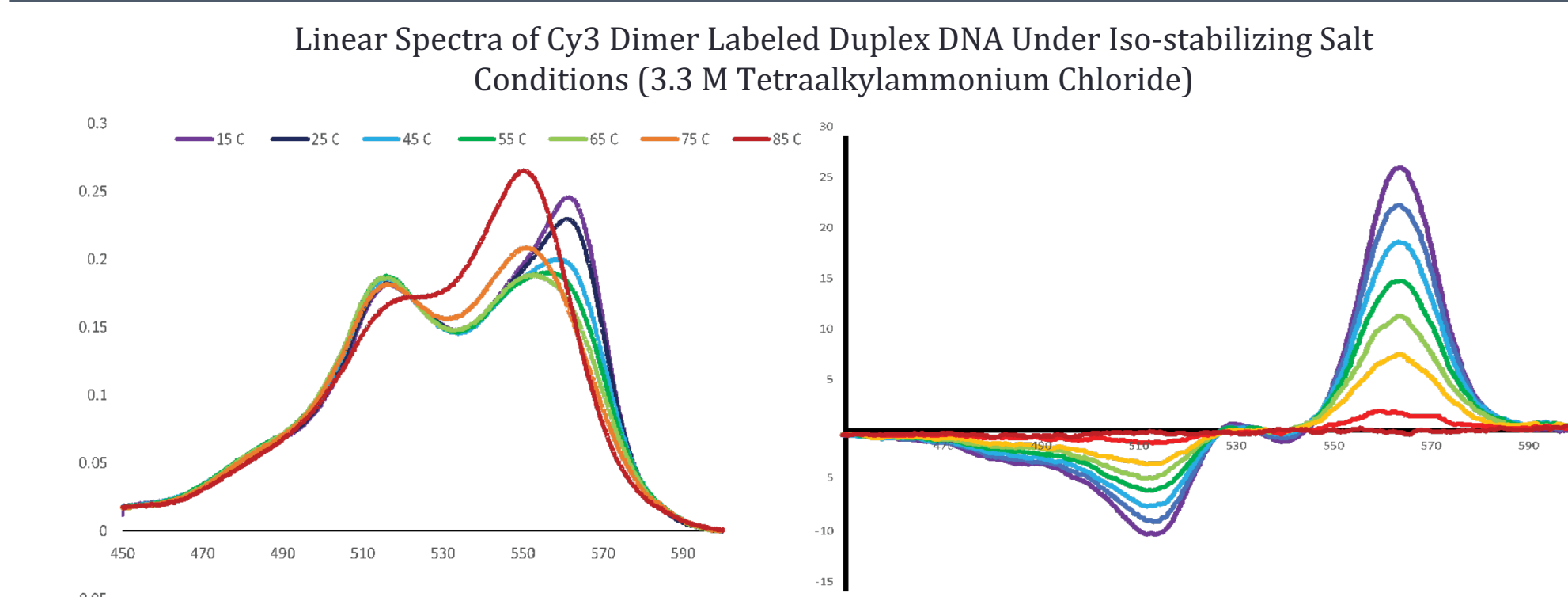
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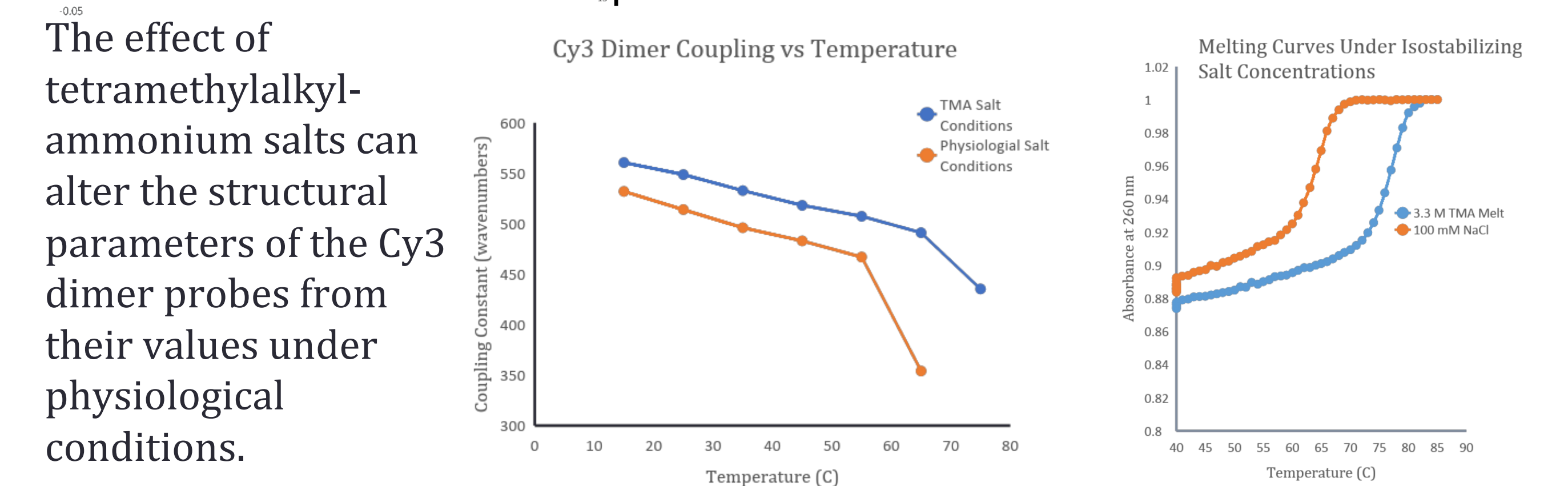


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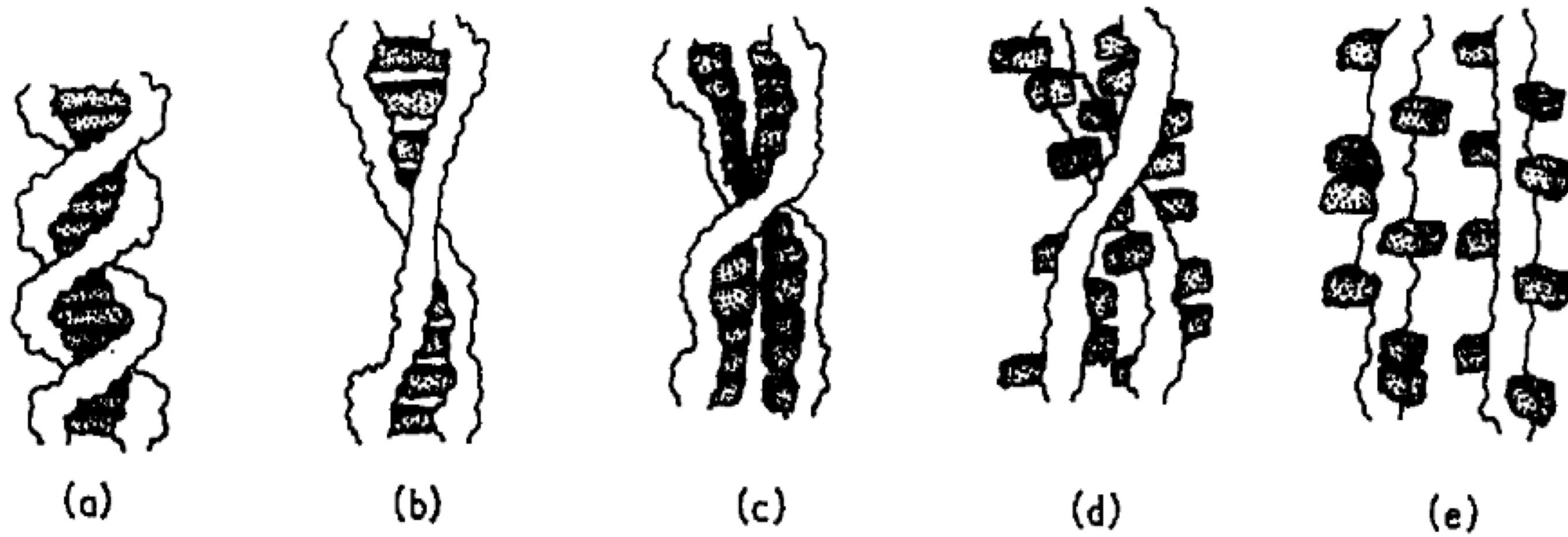


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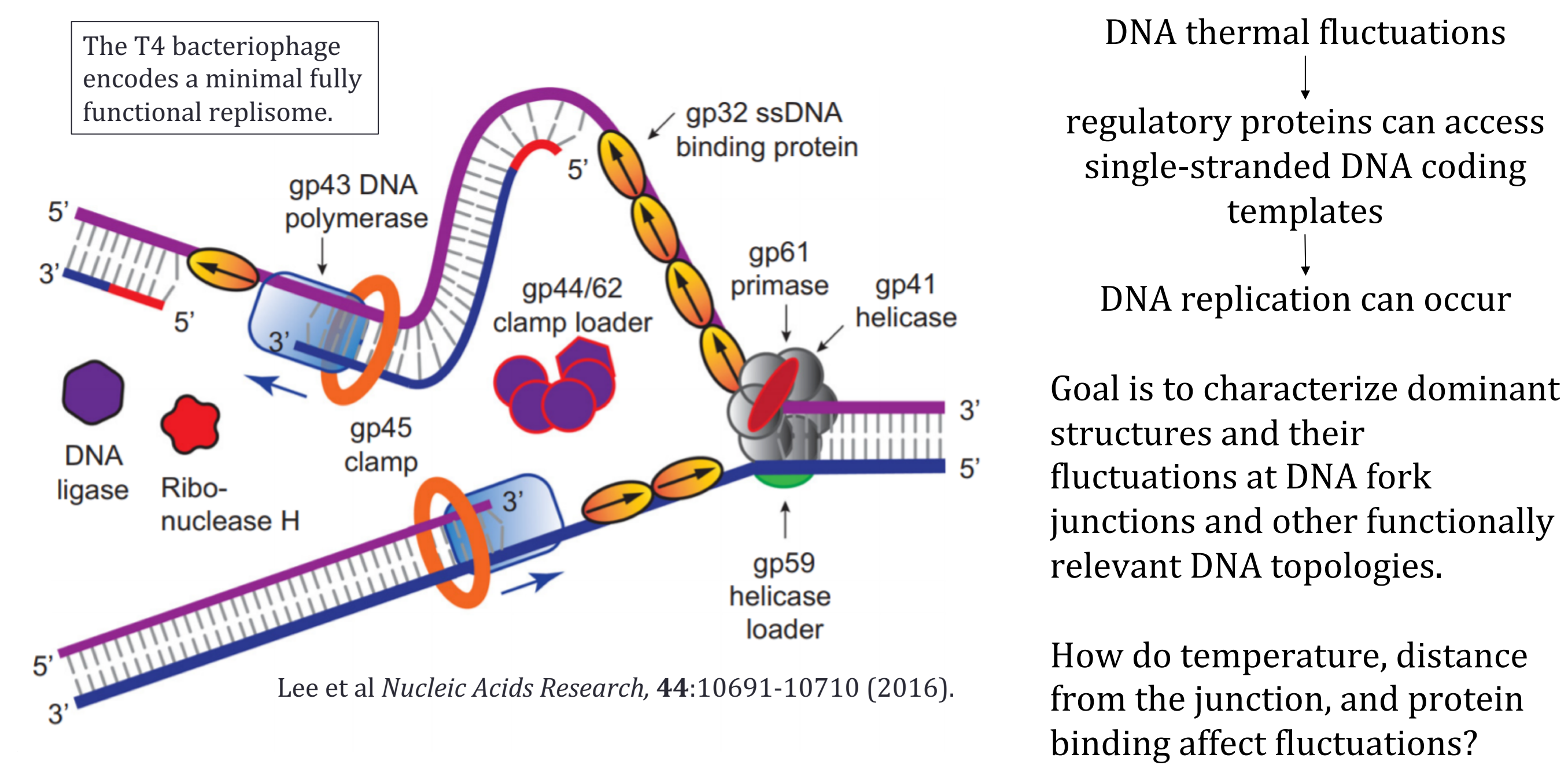
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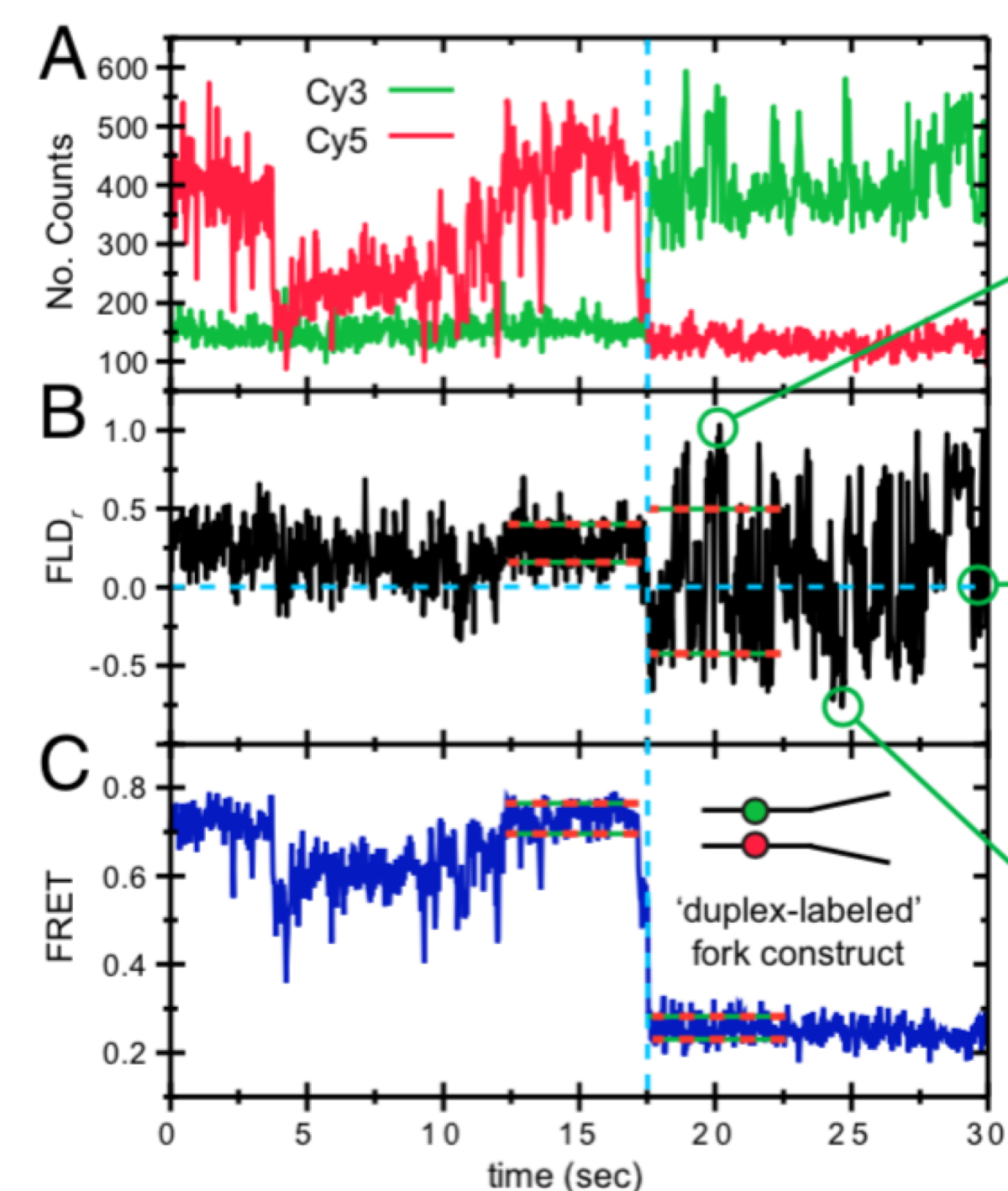
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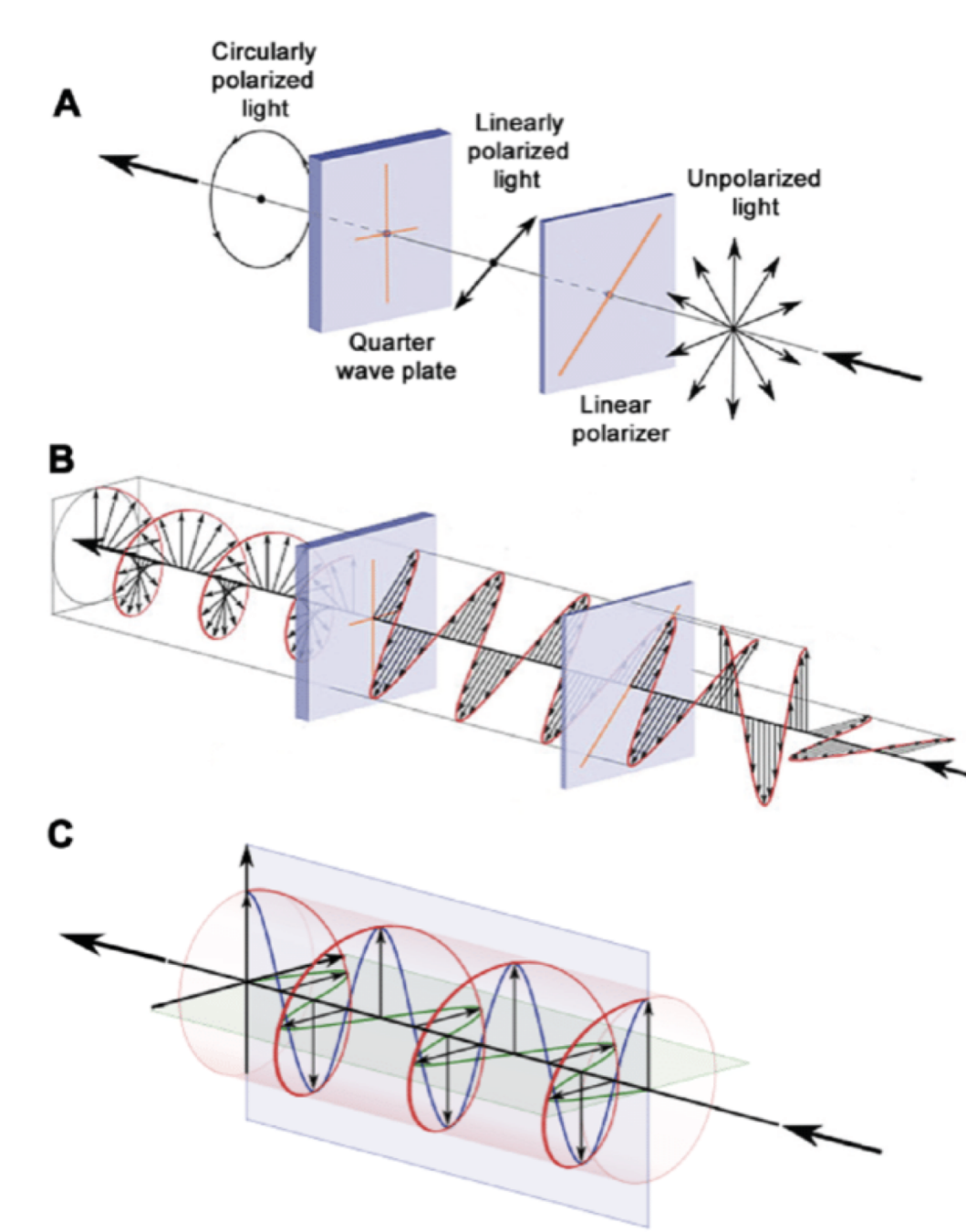
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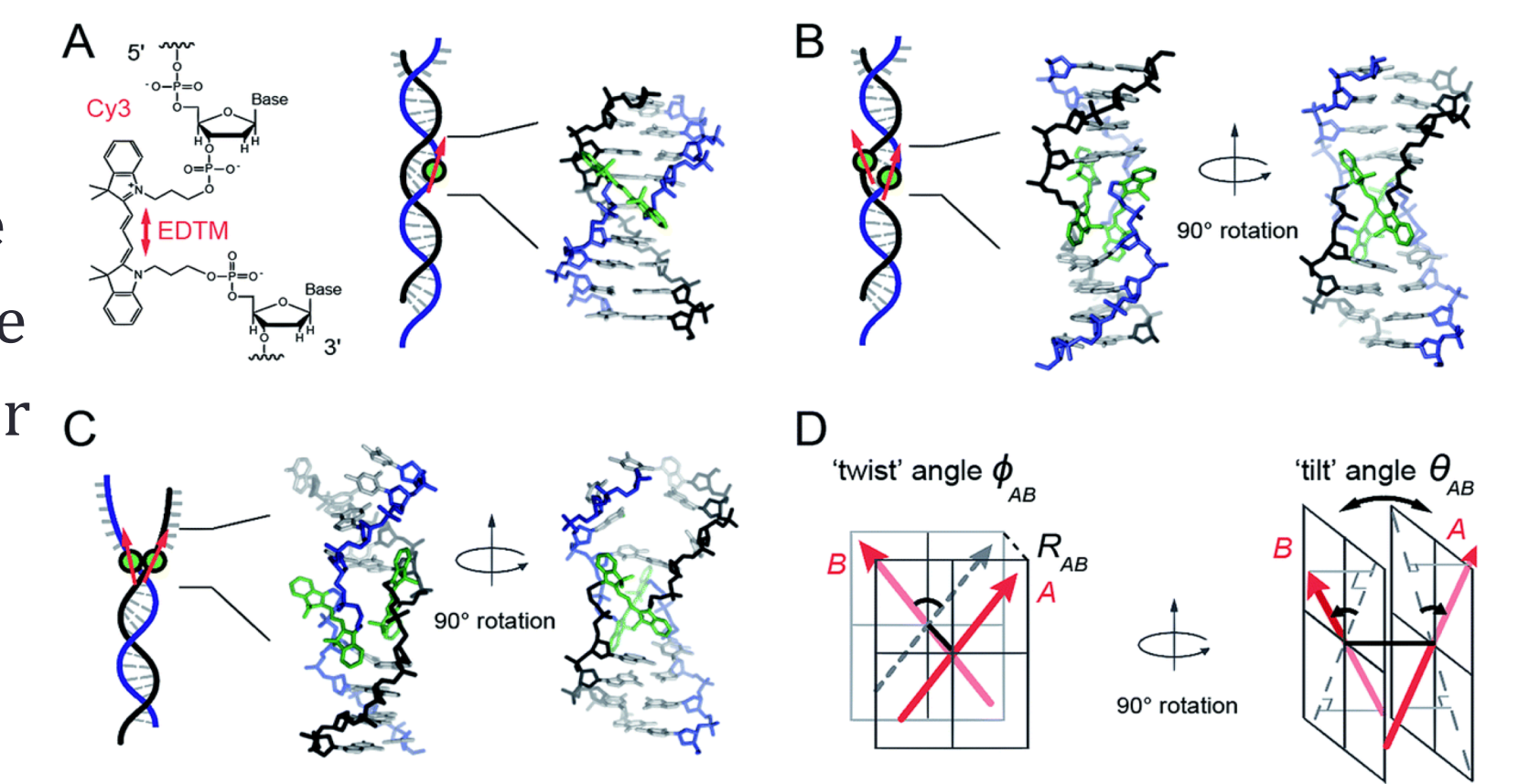
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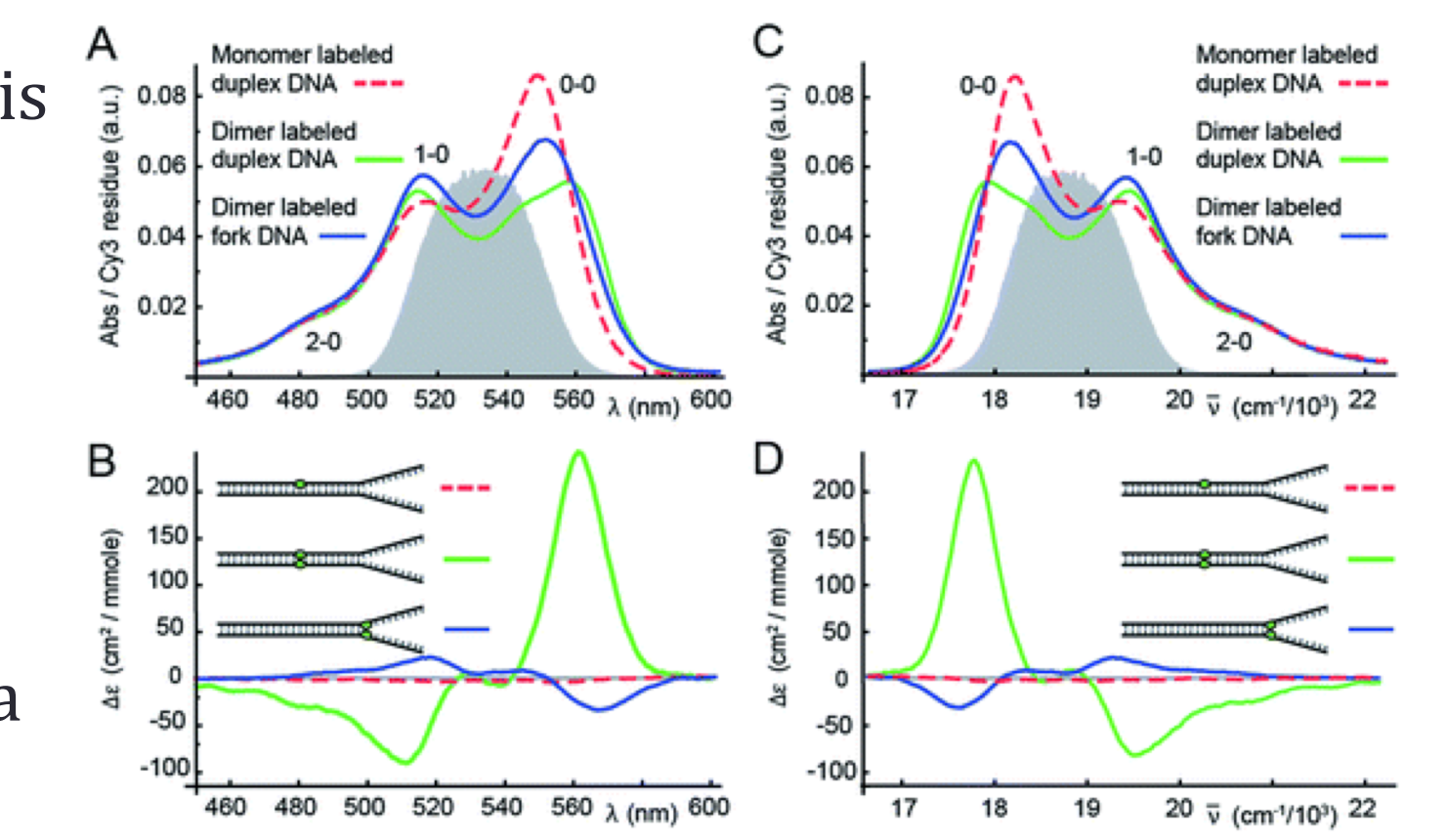
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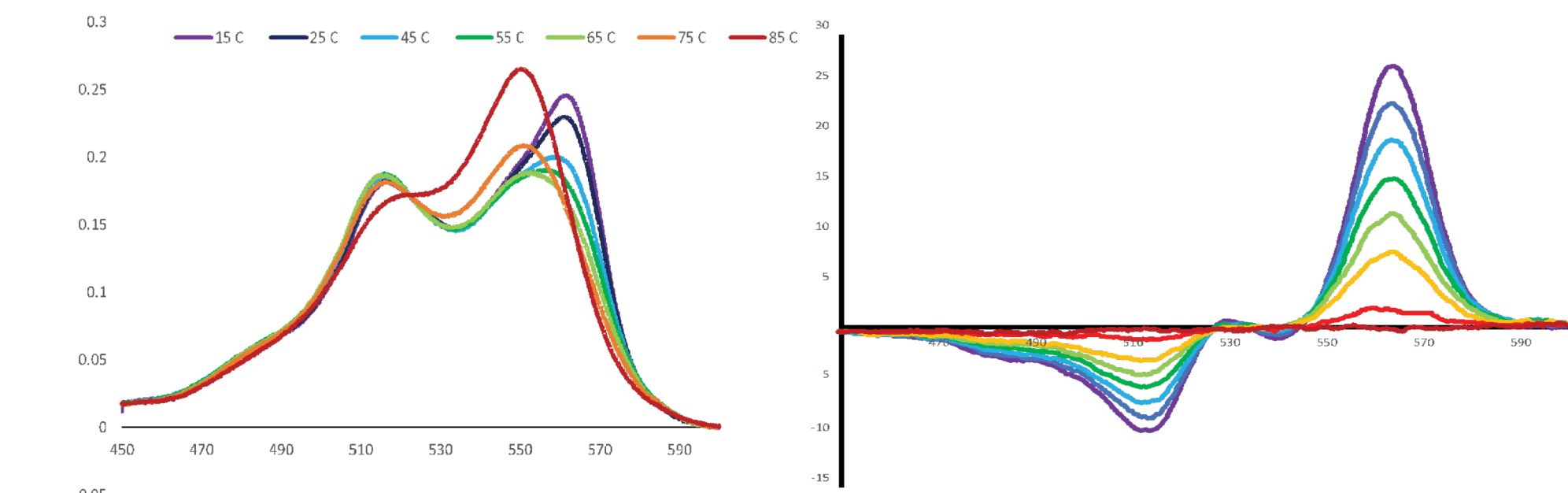
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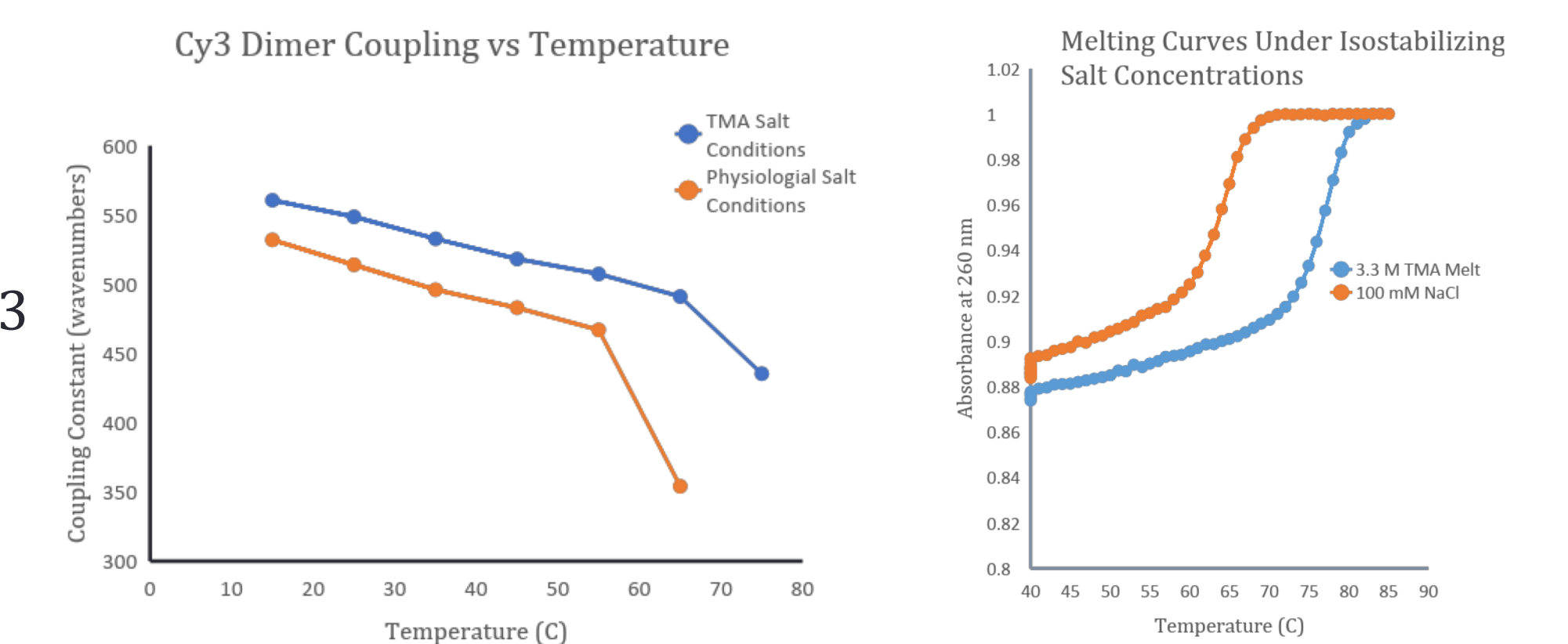
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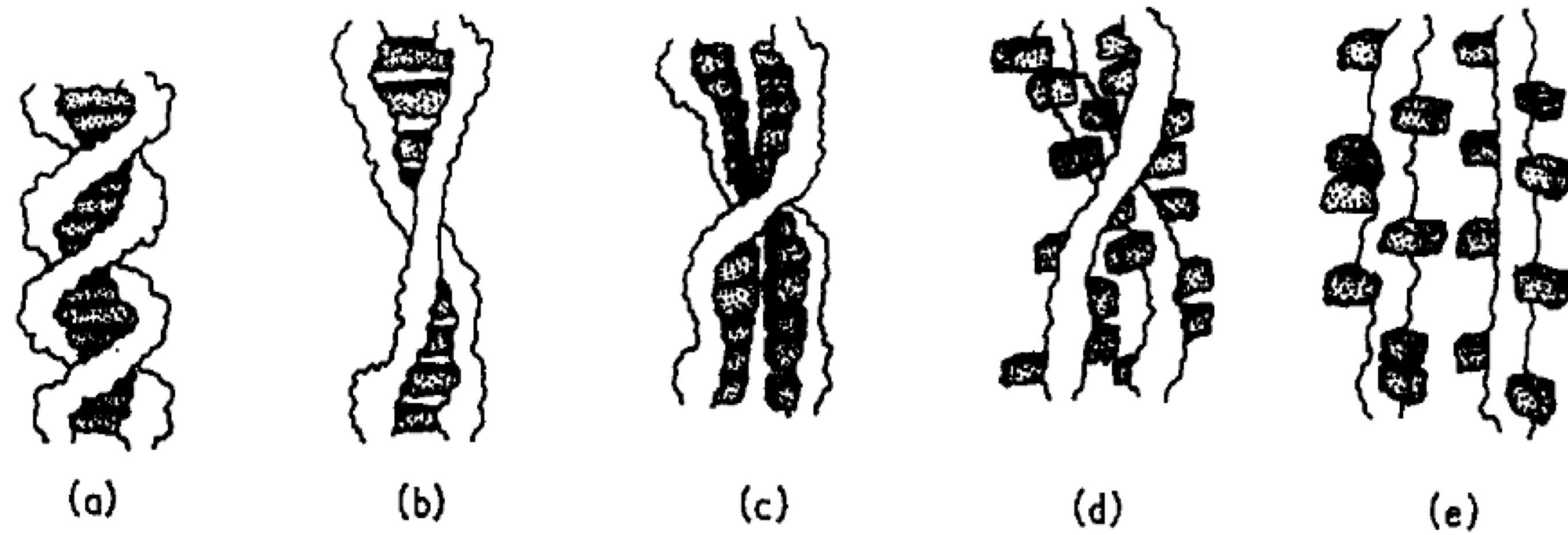
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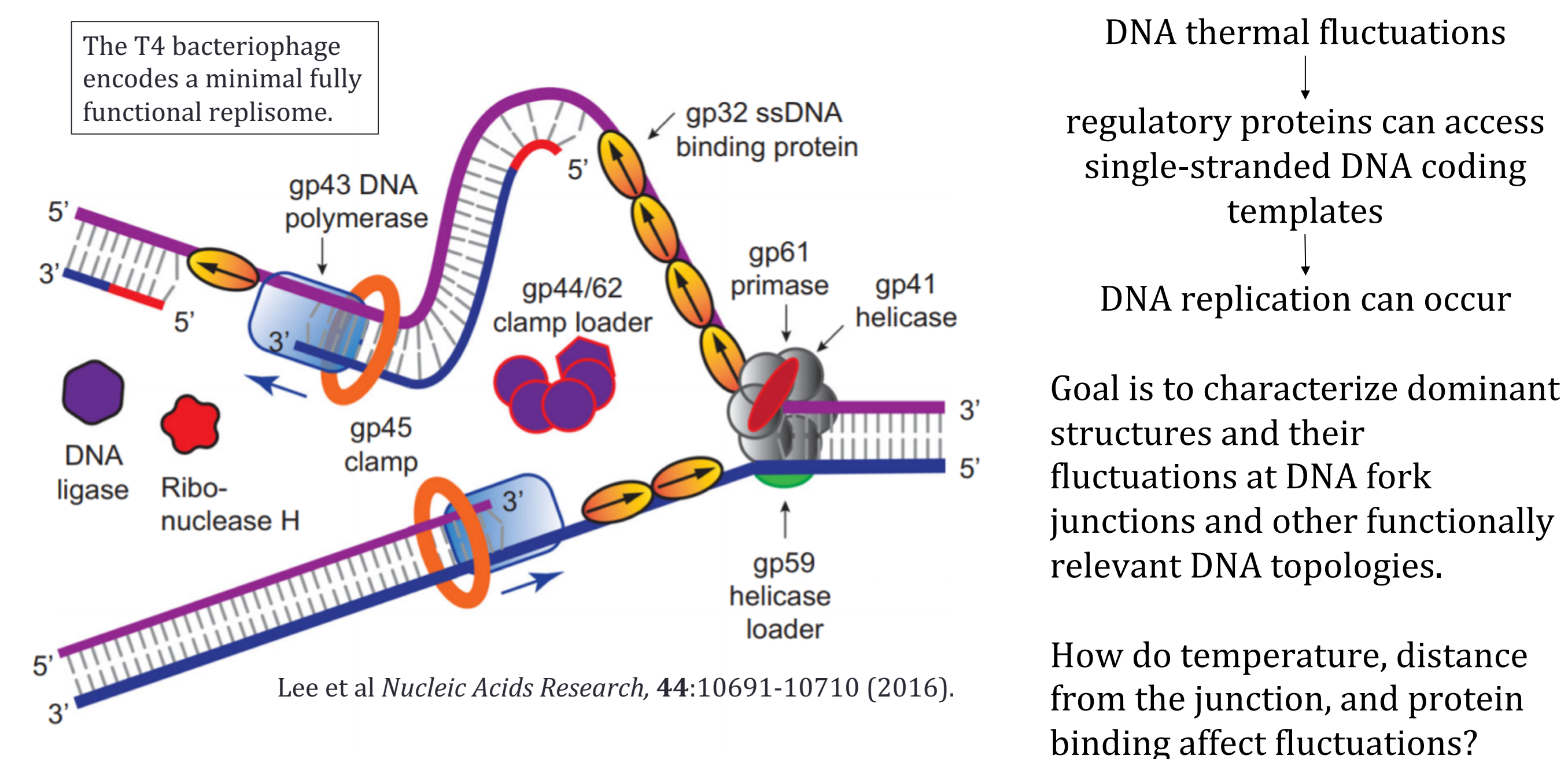
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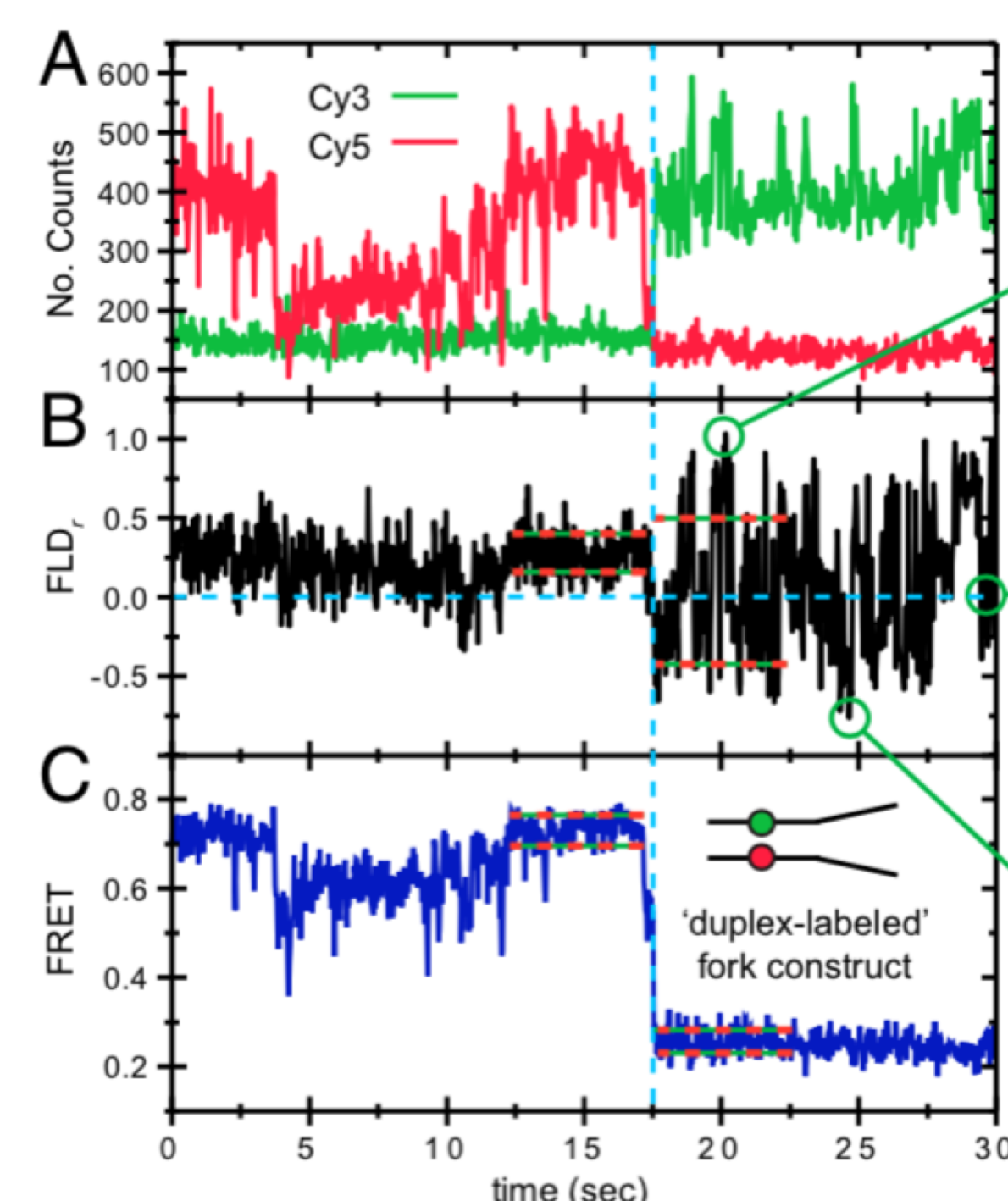
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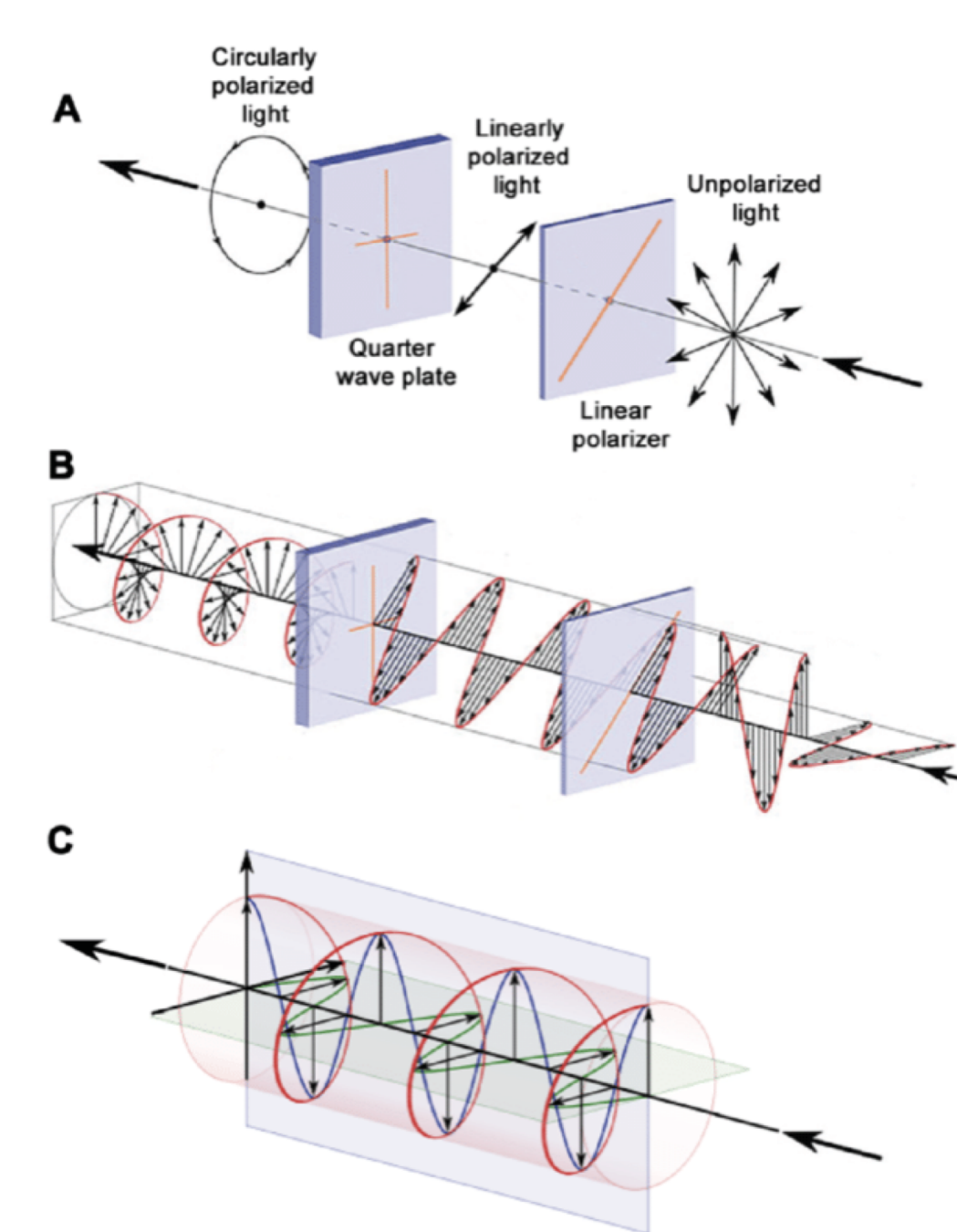
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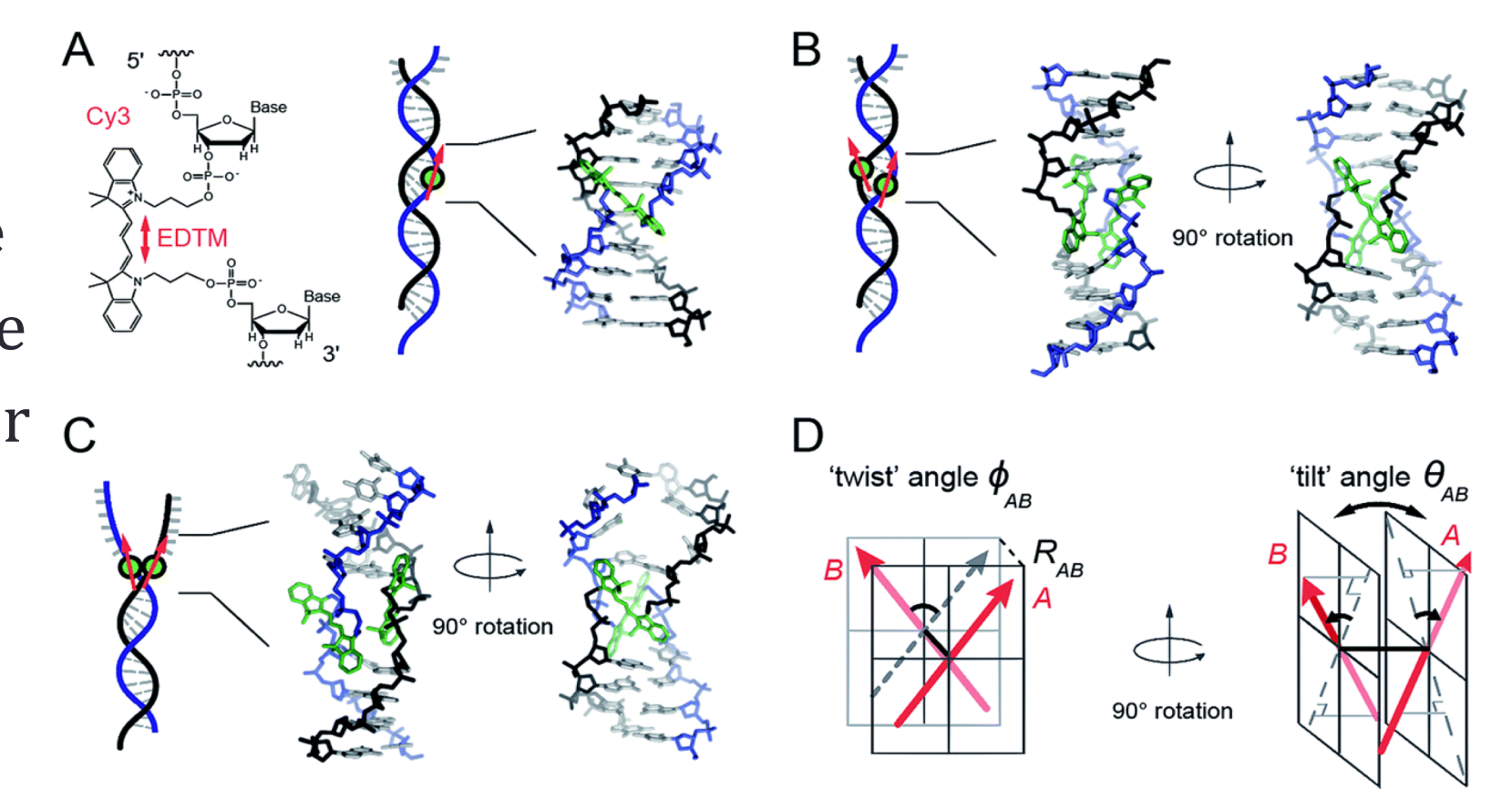
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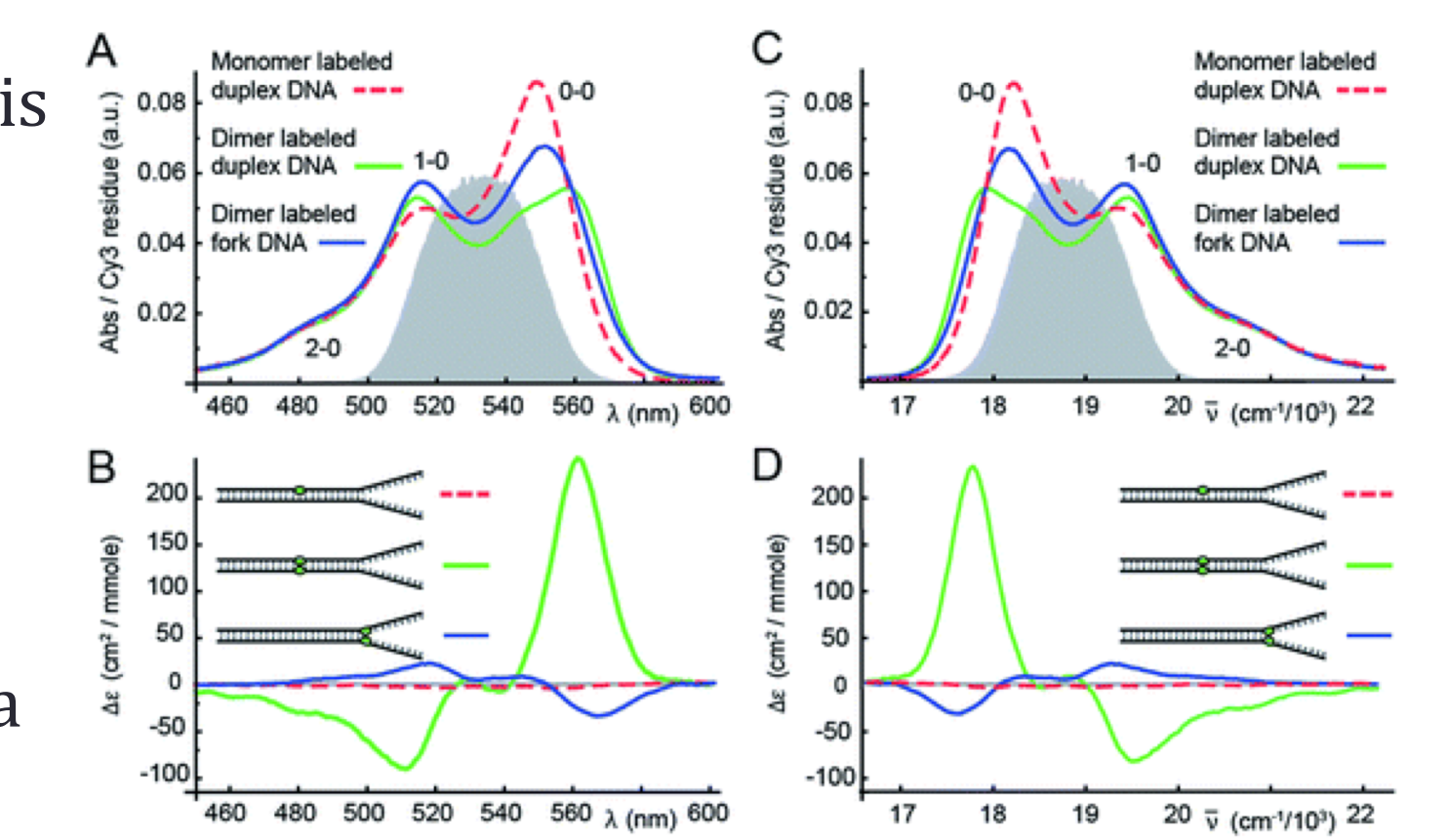


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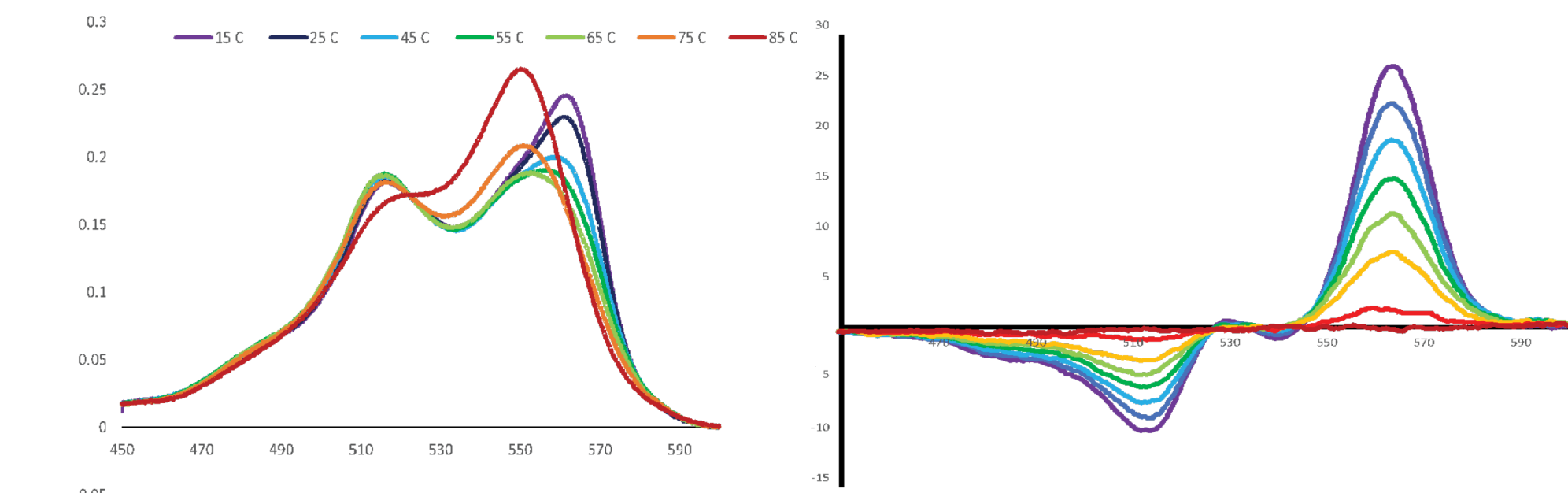
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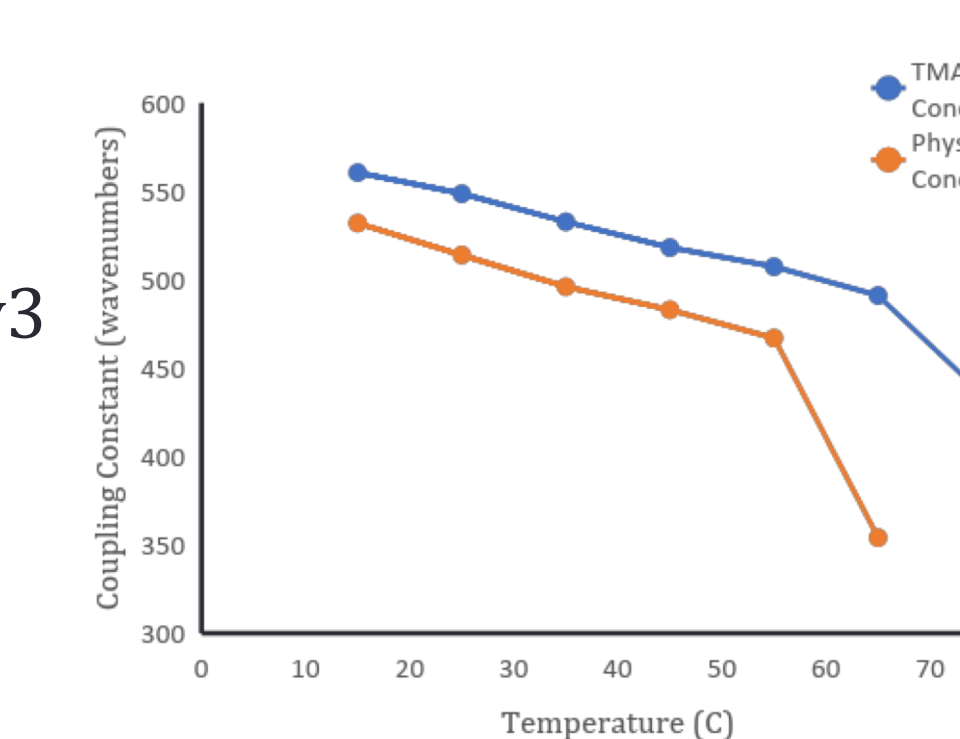
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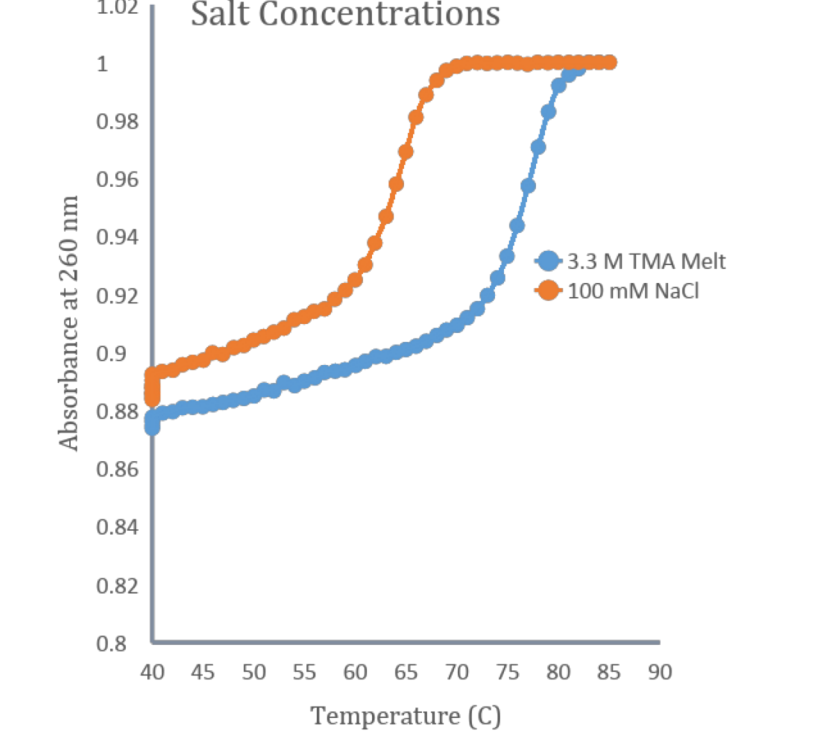
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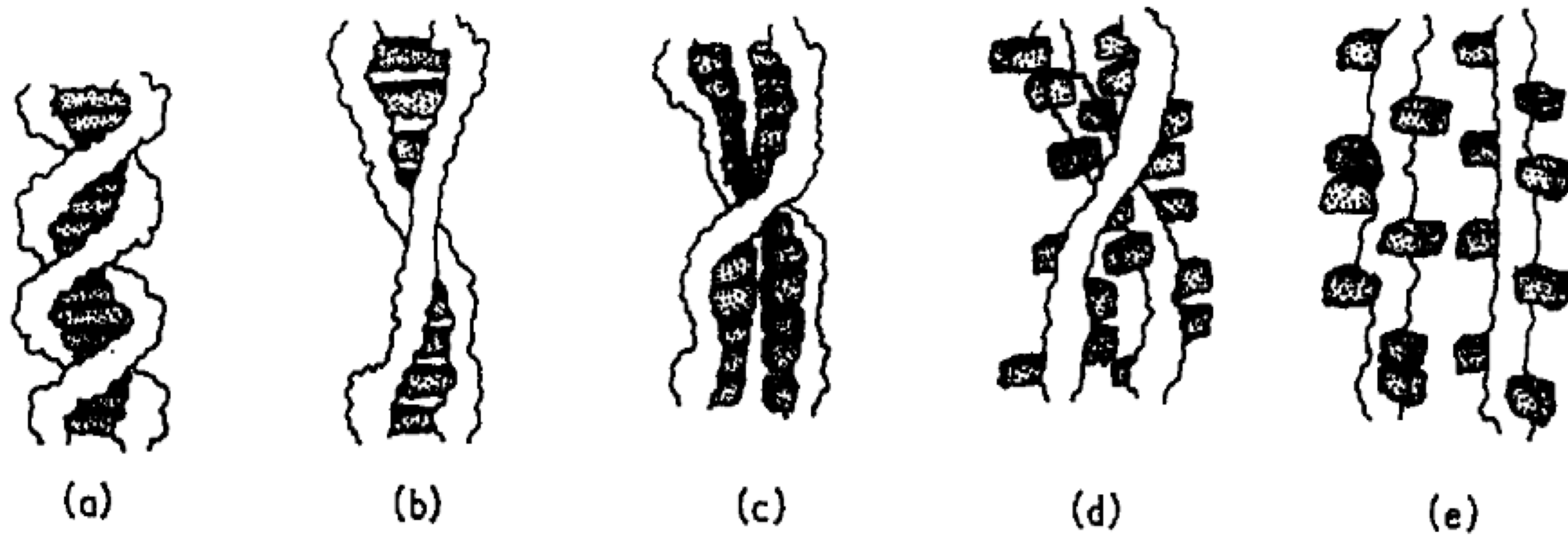


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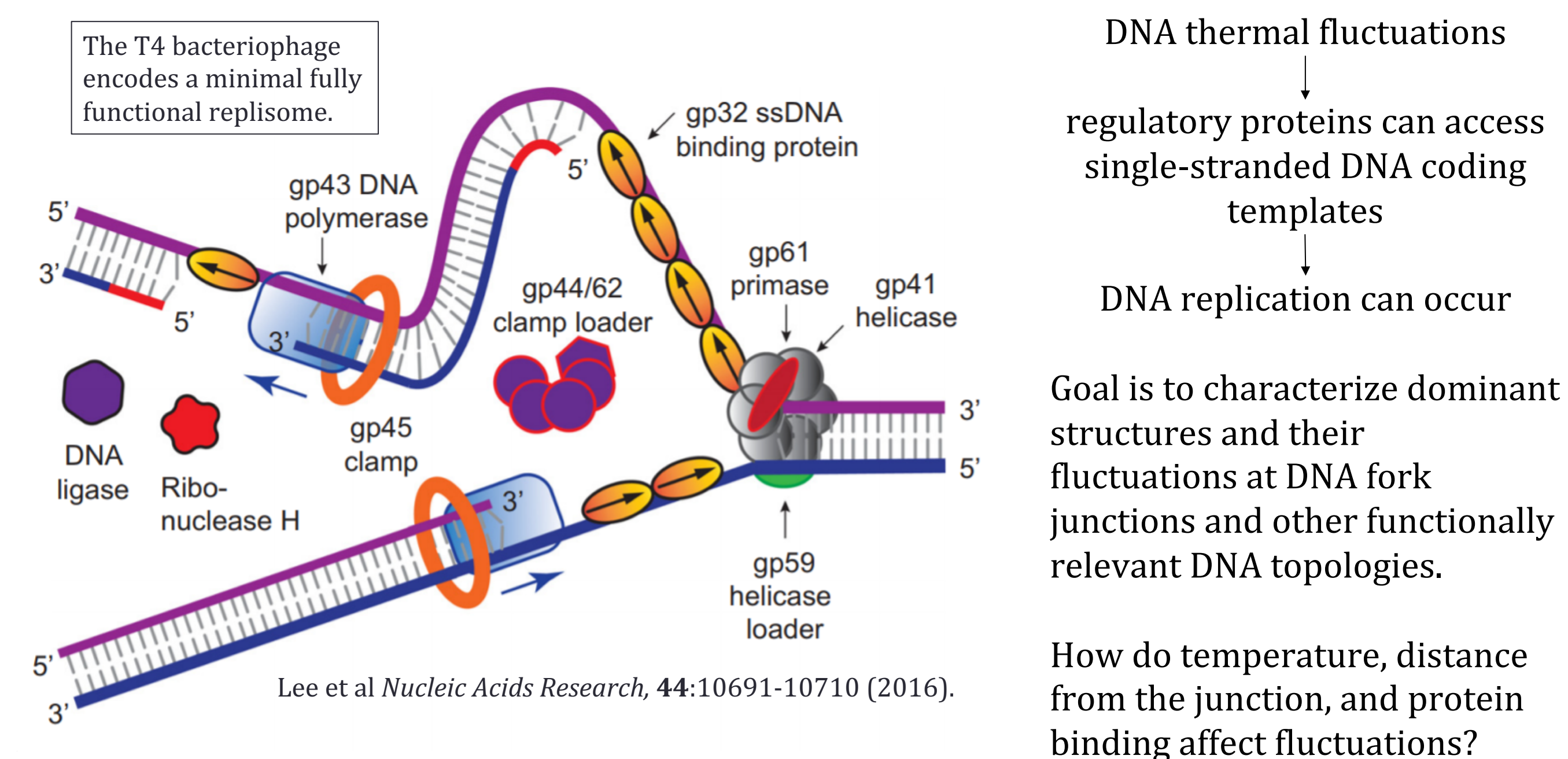
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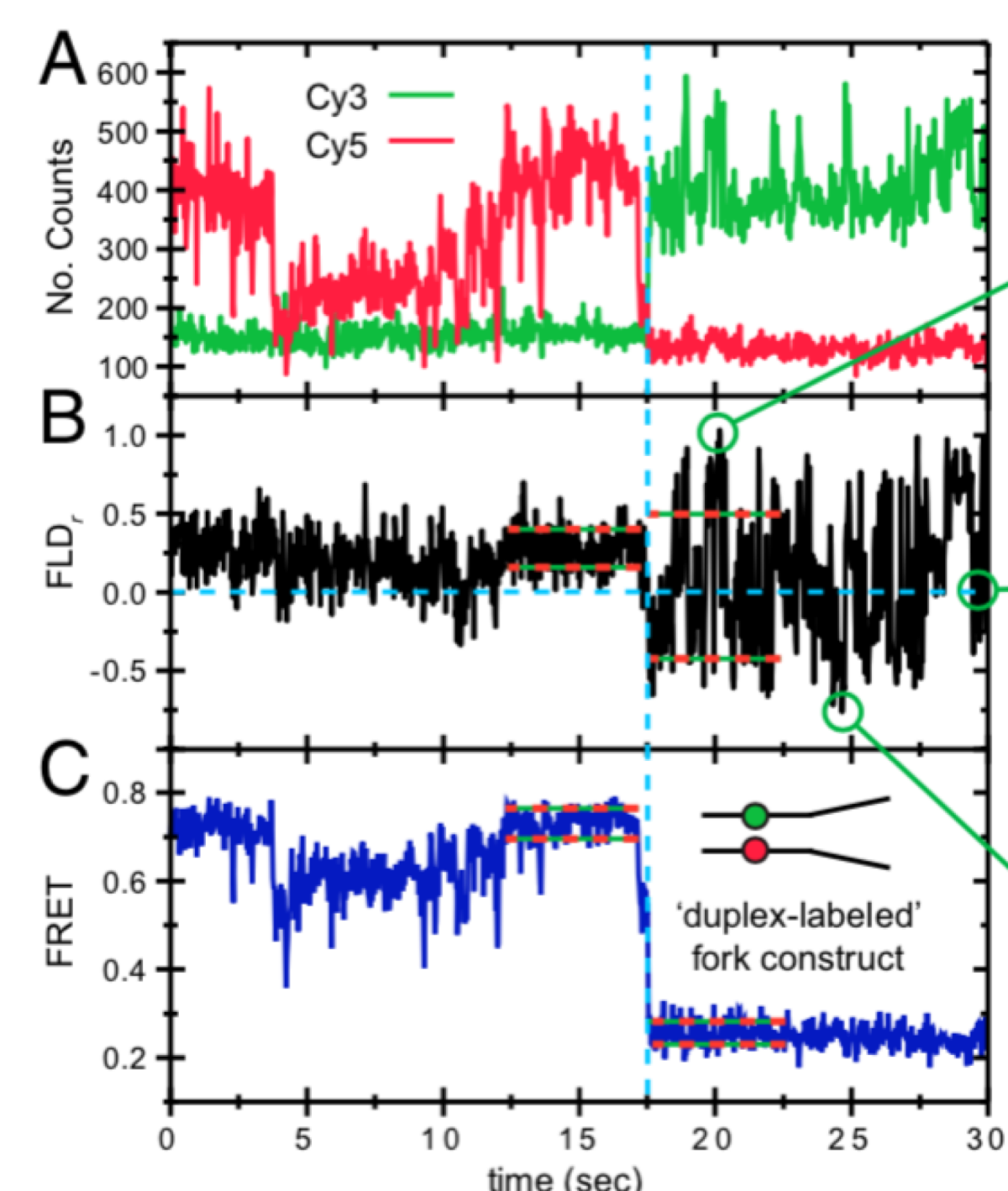
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In our experiments, we used a variety of techniques to analyze the structure of DNA with the overall goal of better understanding the conformations it can take. Our studies were focused in two areas:

1. Understanding the mechanisms of DNA breathing. Duplex and fork DNA undergo thermal fluctuations under physiological conditions that allow for the binding of proteins.
2. Conducting experiments on the stabilizing and destabilizing properties of salt solutions on DNA. We also looked to understand the method by which iso-stabilizing salts stabilize the conformation of DNA even under varied temperatures.

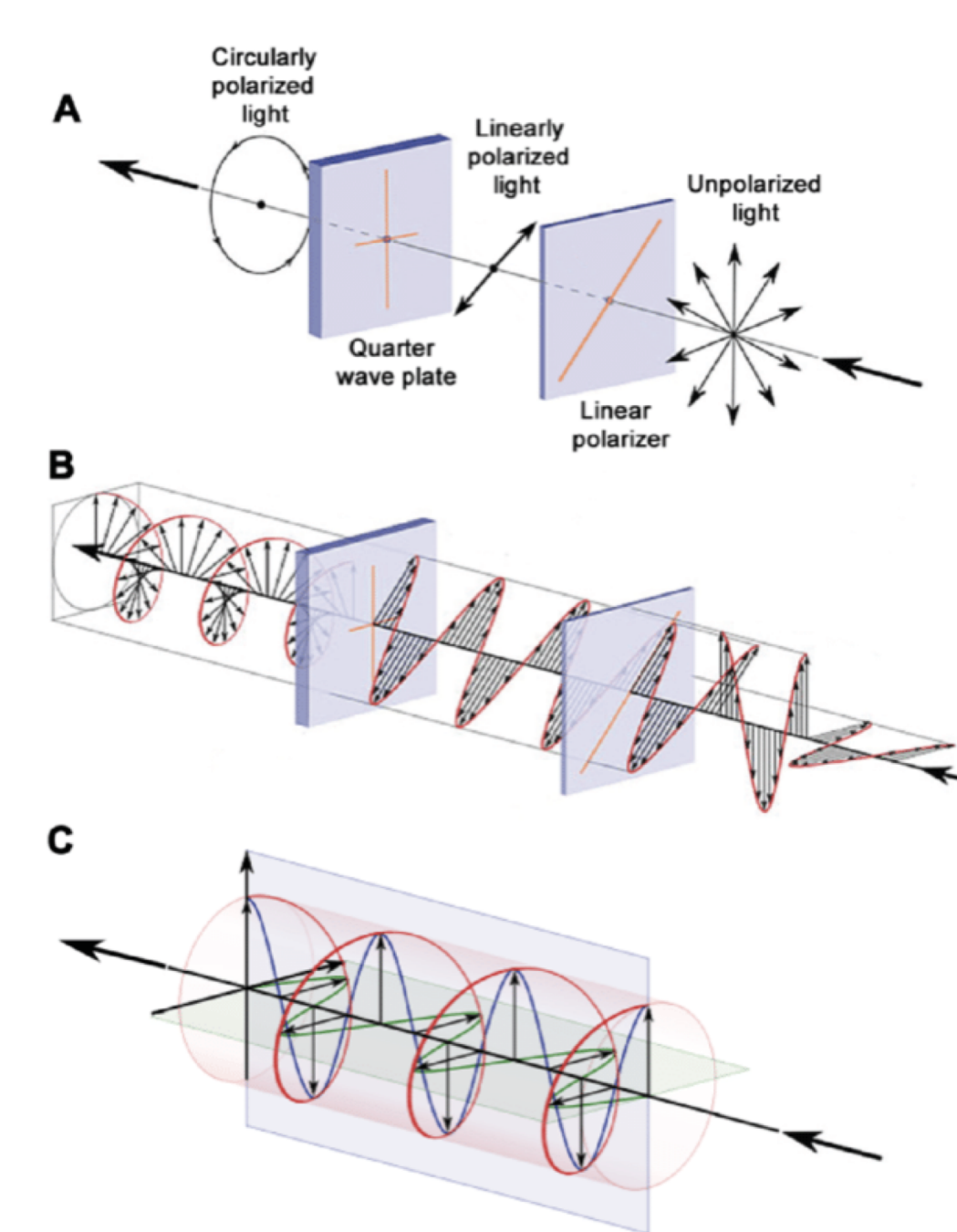
Both studies were done to aid further investigations within the Marcus Group at the University of Oregon.

INTRODUCTION AND BACKGROUND

The Marcus Group conducts studies on the dynamics of macromolecules in biological environments. The group employs a variety of methods in order to better understand biologically relevant molecules, including circular dichroism and single-molecule Förster resonance energy transfer (smFRET).

DNA is internally labeled by replacing specific regions of the sugar-phosphate backbone of the replication fork with a Cy3-Cy5 fluorophore pair. Both single-molecule Förster resonance energy transfer (smFRET) and single-molecule fluorescence-detected linear dichroism (smFLD) are used to study single DNA molecules. Studying fluctuations in smFRET inform about the time scales on which distance and orientation between the fluorophore pair change. Measuring smFLD informs about the Cy3 orientation with respect to the laser's polarization in the laboratory frame.

Circular dichroism (CD) is another method of absorption spectroscopy that



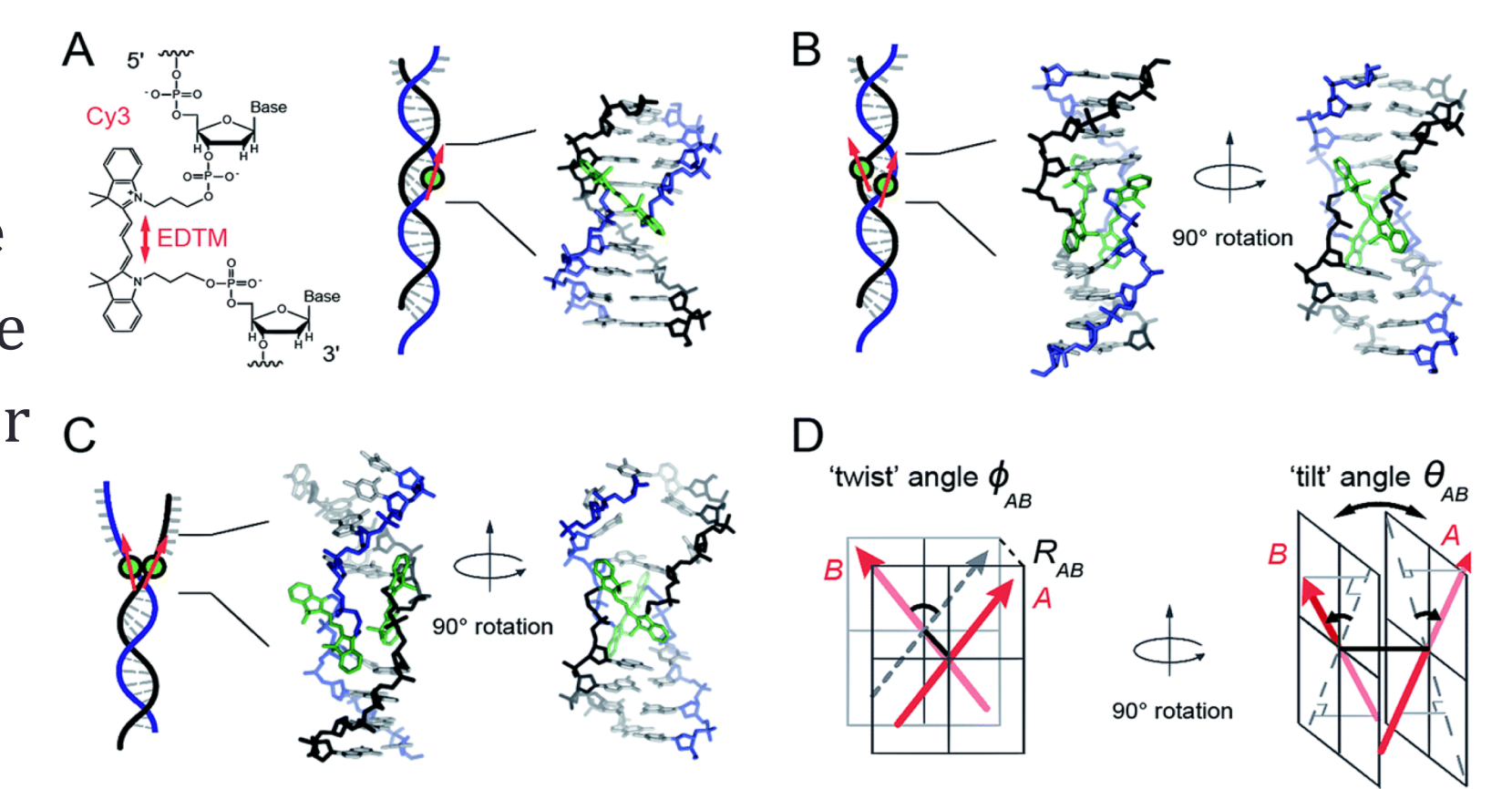
Cronin, TW. 2018, May. A different view: Sensory Drive in the polarized-light realm. *Current Zoology*, 64(4)

utilizes the differential absorptions of chiral light (left and right polarizations). Linearly polarized light occurs when the magnetic and electric fields oscillate perpendicular to one another, forming a wave. In circular dichroism, the electric field remains the same while the magnetic field causes charge to circulate. Circularly polarized light separates into right and left circularly polarized light in an optically active medium. The difference in their absorptions is measured by circular dichroism.

SITE-SPECIFIC CY3 LABELING OF THE SUGAR-PHOSPHATE BACKBONE OF DNA

The sugar-phosphate backbone of DNA is labeled with Cy3

fluorescent dye. Cy3 is site-specifically incorporated into the backbone in order to measure the conformations of the dimer under varying conditions. Cy3 dimer probes "couple" with varying strengths when brought close together. Electronic coupling can be studied through properties of absorption and fluorescence.

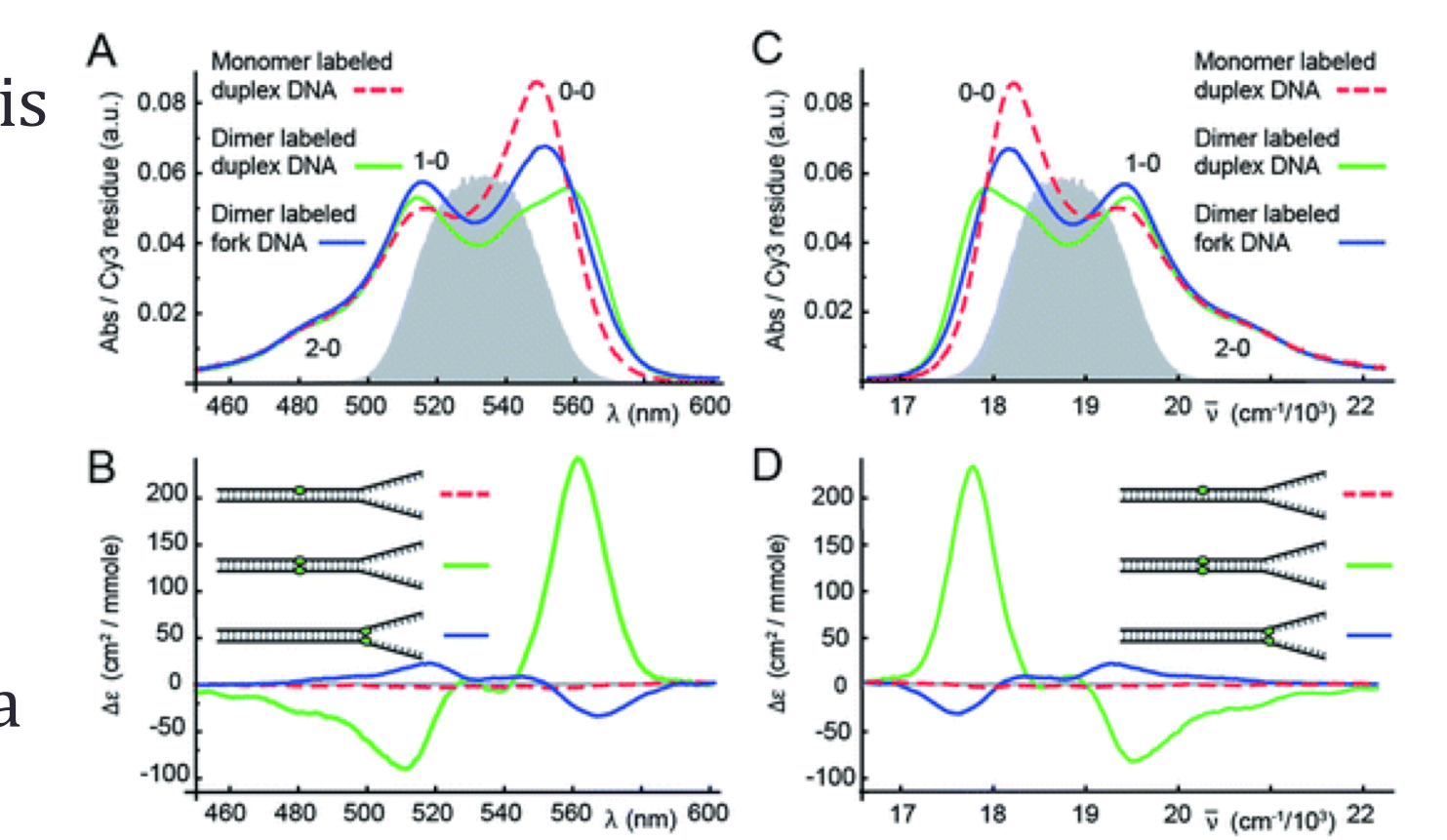


Heussman, Kittell et al. *Faraday Disc.* **216**, 211 (2019).

CIRCULAR DICHROISM AND ABSORBANCE STUDIES OF CY3 LABELED MONOMER AND DIMER DNA

Conformational fluctuations of Cy3 labeled DNA are measured using UV-vis absorption spectroscopy and circular dichroism (CD). The CD of monomer labeled duplex DNA at 25 °C is weak because no coupling can occur. Both dimer labeled constructs exhibited a change of sign within vibronic bands, a signature of vibronic excitations.

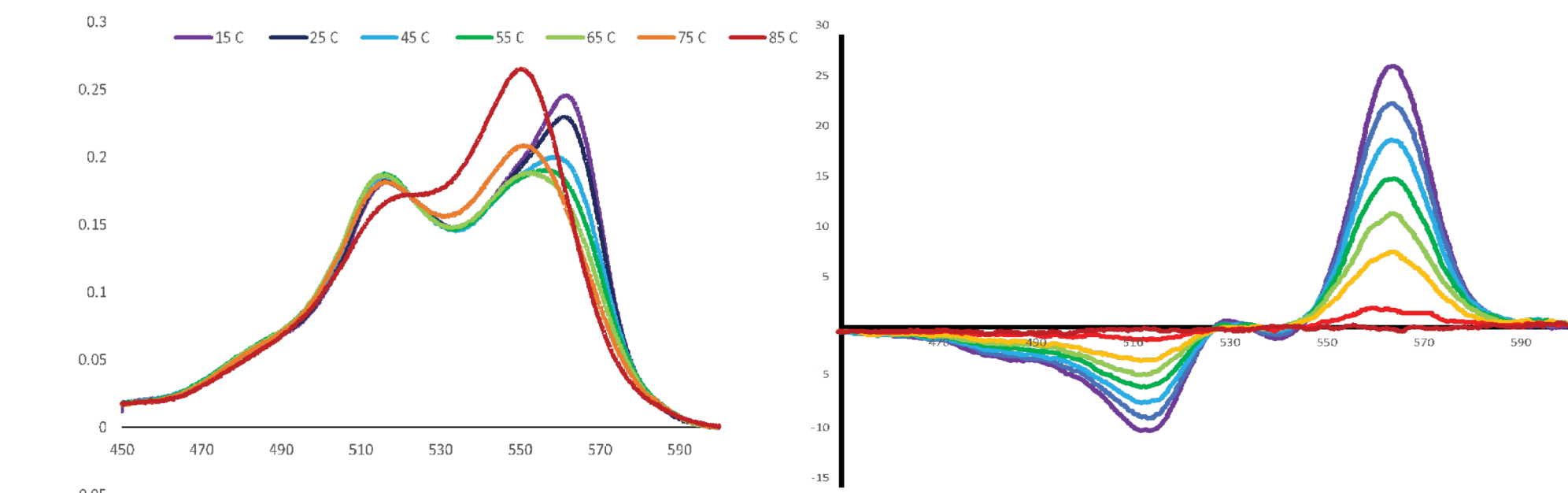
Additionally, CD signals reveal different chiralities of the duplex and fork DNA.



Heussman, Kittell et al. *Faraday Disc.* **216**, 217 (2019).

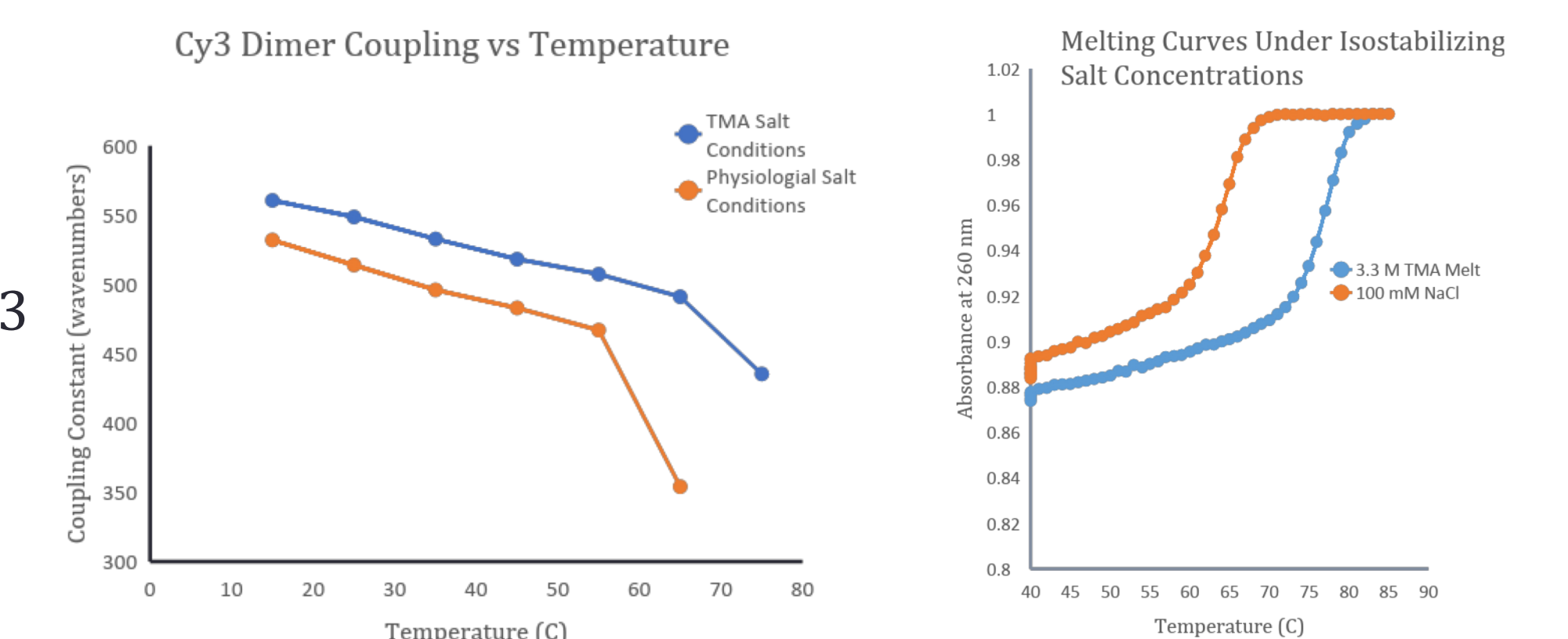
ISO-STABILIZING SALTS MODIFY THE FREE ENERGY LANDSCAPE OF CY3 LABELED DNA

Linear Spectra of Cy3 Dimer Labeled Duplex DNA Under Iso-stabilizing Salt Conditions (3.3 M Tetraalkylammonium Chloride)



The Cy3 dimer probes show the presence of iso-stabilizing salts stabilizes the conformation of the sugar-phosphate backbone.

The effect of tetramethylalkylammonium salts can alter the structural parameters of the Cy3 dimer probes from their values under physiological conditions.



Future studies may focus on fork rather than duplex DNA. Tetramethylalkylammonium is known to raise the melting temperature of DNA. By measuring the CD and absorption over larger temperature ranges or increasing coupling between the two Cy3 labeled sites, we may be able to learn about the structural changes occurring during melting and the stabilizing properties of TMA.