OSMOTIC AND IONIC REGULATION IN GRUSTACEA

by

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Preface

This paper is an introduction to the processes of osmoregulation and ionic regulation in Grustacea, and a selected review of the literature pertinent to a study of those processes. The problems of adaptation to the environment and flexibility in the face of changing environmental influences are basic to an understanding of the evolution of morphological and physiological characteristics. This paper will attempt to elucidate the role of regulation of ions and water in facilitating ecological flexibility, and the limits imposed on creatures that lack this ability.

* * *

Every animal is inescapably in some form of contact with his environment, and the interaction between that environment and the animal's body fluids is a matter of crucial importance. Life began in water, and as living creatures are composed, on the average, of 70-85% water (Nicol, 1960), direct exposure to air calls for relatively sophisticated mechanisms to avoid partial dessication. Terrestrial animals must therefore be equipped with a means to conserve water, and even marine forms often must work to retain water although they actually live immersed in it. If the concentration of ions in the body fluids of a marine animal is less than the concentration of ions in sea water, water may move out of the animal by osmosis. The result may be that the animal is in need of water to dilute its body fluids to a concentration which will allow normal physiological processes to continue. Conversely, fresh-water animals have the problem of maintaining a finite amount of water in their bodies. Since the medium is less concentrated than their body fluids, ions may diffuse out into the medium, and water may enter such animals by osmosis. They then have the problem of preventing too great a dilution of their internal ionic concentration. Thus every animal, no matter where it lives, must cope with the stresses of a basically hostile world.

It is not enough to consider the problem solved if an animal does maintain an optimum amount of water in its body; that is, the entire problem is not to achieve a set concentration. Certain ions are required, and the body must have them in definite proportions. If the external medium does not offer such ions in the concentrations necessary, an animal must find some means to selectively withdraw them from the medium; without such mechanisms the animal must assume the concentration for each ion as it is represented in the medium, and often this value is not within the limits set for an optimum balance of chemicals.

Some Useful Terms

Every water medium has a certain salinity, or concentration of salt, which may be expressed in parts per thousand (%o). Thus sea water has a salinity of approximately 34.5 %o, while fresh water has a salinity of less than .5%o (Prosser and Brown, 1950). Physiologists label animals which are able to cope with a wide range of environmental salinities as euryhaline, and those which can exist successfully in only a narrow range as stenohaline. Many animals passively adopt the osmotic concentration of the medium. Such animals are termed osmoconformers, or polkilosmotic; if, instead, an animal maintains a fairly constant internal concentration, despite the concentration of the medium, it is termed an osmoregulator, or homoiosmotic.

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Osmoregulation refers to regulation of the total particle concentration of the body fluids at levels which may differ from the concentration of the medium. A solution which is more dilute than another is hyposmotic relative to that solution; if it is more concentrated, it is hyperosmotic. When, instead, water is balanced between two systems, they are isosmotic.

Ionic regulation is "the maintenance in a body fluid of concentrations of ions differing from those in passive equilibrium with the external medium." (Robertson, 1953). It is useful to define regulation, either osmotic or ionic, as an active, energy-requiring process. At the expense of such energy a successful osmotic and ionic regulator is expected to be able to survive in salinities far different from its internal milieu, while osmoconformers and ion conformers can never leave the medium which approaches their own optimum concentration of ions.

General Principles

A membrane which is permeable to water molecules but not to solute particles is termed a semipermeable membrane. If such a membrane separates two solutions of different concentrations, water will move into the more concentrated solution until the two solutions have the same molal concentration.(Molal = moles of solute/kg water.) This movement of water is osmosis (Potts and Parry, 1964). An equation for osmosis of water into a body of concentration C2 from a medium of lower concentration C1 is given by $v_w = k_w$ (C2-C1), where v_w is the rate of water movement and k_w is a rate constant (the permeability coefficient of the external body surface to the water) (Croghan, 1961).

Diffusion is the process of random mixing of solute particles in a solution of unequal particle distribution until an even distribution is reached. There is a net movement of particles from a place of greater concentration to a place of lesser concentration (Potts and Parry, 1964). Two solutions, of concentrations C1 and C2, separated by

a semipermeable membrane of thickness T, constitute a concentration gradient of C2-C1/T. The rate of diffusion across the membrane in an area A is DA C2-C1/T, where D is a diffusion constant (Potts and Parry, 1964). For a body in a medium less concentrated than itself, the equation for solute lost from the body by diffusion is $r_d = k_g$ (C2-C1), where r_d is the rate of solute diffusion, k_s is a rate constant (the permeability coefficient of the external surface to the solute), C2 is the body fluid concentration, and C1 is the medium concentration (Croghan, 1961).

If there is a potential difference between two solutions due to the net charge of their respective solute particles, then there is a potential gradient between the solutions, and charged particles will tend to move down the gradient and thereby decrease the potential difference. A potential gradient may either reinforce or inhibit diffusion due to a concentration gradient, depending upon the direction of the gradients.

The Energetics of Osmotic and Ionic Regulation

When molecules move down a concentration gradient, energy is liberated. To move a solute against a gradient requires energy. The work, W, required to move 1 mole of solute from a solution of molal concentration C1 to a solution of higher concentration C2 is W = R T ln (C2/C1)

(Potts and Parry, 1964). If there is also a potential difference of E volts between the solutions, there is an energy requirement of zEF to move 1 mole of ion against the electrochemical gradient, where z is the valency of the ion and F the Faraday. Then the equation for work is $W = R T \ln (C1/C2) + zEF$ (Potts and Parry, 1964).

If uptake proceeds from a concentration C1 to a concentration C2 by carrier sites with a high affinity for solute particles in C1, the rate of transport from C1 to C2 is determined by the number of sites and is independent of the concentration of solute in C1. (The carrier site is saturated at very low concentrations of solute in C1.) Croghan (1961) says that if one assumes that the activation energy of transport is high, then the system must operate well away from the equilibrium position to maintain an appreciable rate. Therefore, he states, the transport system is probably unidirectional and the rate of transport by a given number of sites is, for the most part, independent of the body fluid concentration. Then rm, the rate of active uptake from the medium, = $k_m n_m$, and r_t , the rate of active uptake from the excretory tubule, = $k_t n_t$, where k_m and k_t are the rate constants of the carrier mechanisms and n and nt are the numbers of active carrier sites (Croghan, 1961). Shaw

(1958), in a study of Na absorption in the crayfish Astacus ballipes, verified that in a medium of low concentration the rate of active uptake is proportional to the concentration of the medium. The concentration determines the number of ions coming into contact with transport sites. At high concentrations of the medium all transport sites are saturated, and absorption of ions is independent of the concentration. Given the same permeability and blood concentration, the energy cost of osmoregulation is then dependent on the surface to volume ratios of the animal. A small animal will require more work per unit mass of body tissue than a large animal to maintain a certain blood concentration. A decreased blood/medium concentration gradient will lessen the work, and a smaller animal usually maintains for this reason a lower blood concentration than does a larger animal living in the same medium (Lockwood, 1962).

When water is balanced, $v_w = v_u$, where v_u is the rate of urine production. Solute will then enter the excretory organ at a rate, v_f , = k_w C2 (C2-C1) and solute will be lost from the animal via the urine at a rate $v_u = k_w C_u$ (C2-C1) where C_u is the concentration of the urine excreted (Groghan, 1961). The rate of active uptake from the excretory tubule is $r_t = k_w$ (C2-C_u)(C2-C1). The rate of active uptake from the medium is $r_m = k_g$ (C2-C1) + $k_w C_u$ (C2-C1). When C1 is

small compared to C2, C2 = $\forall r_m + r_t / k_w$ (Croghan, 1961). Therefore the concentration of body fluid in a given medium concentration is determined by the permeability coefficients and the rates of the uptake mechanisms.

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Fresh-water Crustacea

Crustaceans in fresh water must keep their body fluids hyper-osmotic to the medium because they would have body fluids too dilute for cellular functions if they were isosmotic to the fresh water. Water is continually entering osmotically, and ions leaving their bodies through excretion and diffusion. Because it requires energy to maintain an internal concentration against such a gradient, fresh-water animals have found it useful to acquire a toleration to a less concentrated internal milieu than many marine organisms. The latter are often isosmotic to sea water and do not have as strong a gradient (Robertson, 1960). Since fresh water does not have the same proportion of ions as required in body fluids, fresh-water forms must regulate ionically as well as osmotically.

To maintain themselves in fresh water, crustaceans may use essentially three mechanisms: 1) a low permeability to water and salts, 2) a selective uptake of ions from the medium at the body surface or from the gut, and reabsorption from the urine, and 3) elimination of water by production of a dilute urine.

The amphipod genus Gammarus has representatives living in a wide range of environmental conditions. Gammarus pulex is a fresh-water form (Shaw, 1961c). It has been suggested that its mechanisms for regulation may in fact be characteristic of fresh-water Crustacea in general. The mechanism it uses for ionic uptake has a high affinity for Na ions, and maximum transport is reached at external concentrations of near 1mM/1. Furthermore, the rate of uptake is increased by only a small drop in the internal Na concentration. The adaptive significance of these facts is better appreciated by comparing this organism with its brackish-water relative, Gammarus duebeni. G. duebeni presents a low affinity for Na ions, and its mechanism does not reach maximum transport of ions until the external concentration is about 10 mM/1. At a medium concentration of .1mM/1, for instance, G. pulex would be capable of transporting about three times as many ions as transported by G. duebeni at the same external concentration. It is clearly a better regulator than G. duebeni in mediums of very low concentrations.

Werntz (1963) showed that activation of the regulatory mechanism in gammarids is gradual, with the critical concentration (the concentration below which maximum regulation rate is employed) being higher in marine species than in fresh-water species. There has been some disagree-

ment as to whether the degree of activation actually depends on the concentration of the external medium or of the blood, but Shaw (1959) favors the former.

Osmotic regulation in four species of gammarids was compared by Werntz (1963). He found that the freshwater species <u>G</u>. <u>fasciatus</u> was able to regulate its blood concentration at a very low level of external salinities, while the brackish and marine forms were able to regulate only at intermediate and high salinities, respectively. Measured by freezing point depression, <u>G</u>. <u>fasciatus</u> maintained a blood concentration of 0.35-0.50 molal% in equivilants of ideal solute, the brackish-water <u>G</u>. <u>tigrinus</u> kept its blood at 0.55-0.65 molal%, and the marine forms <u>G</u>. <u>oceanicus</u> and <u>Marinogammarus finmarchicus</u> maintained concentrations of 0.65-0.85 molal%. This data indicates the toleration by fresh-water forms of more dilute body fluids than maintained by animals living in waters of higher salinity.

There is also evidence in the gammarids for a decreased permeability in the fresh-water animals relative to animals living in more saline waters. Werntz (1963) found that the rate of urine flow in these animals is directly proportional to the osmotic gradient between the blood and the medium, and it is therefore evident that osmotically absorbed water is eliminated as urine. The rate of flow of urine in the

marine <u>G</u>. <u>oceanicus</u> is 10.5% (% of body weight per hour per molal gradient), but in <u>G</u>. <u>fasciatus</u> it is only 5.9%. It is clear that <u>G</u>. <u>fasciatus</u> is less permeable to water than is <u>G</u>. <u>oceanicus</u>. Magnin (1964) has verified that decreased permeability is correlated with adaptation to fresh water.

<u>G. fasciatus</u> reduces salt loss, not only by having a reduced permeability, but also by producing a urine that is hypotonic to its blood. <u>G. oceanicus</u> produces urine that is isotonic to its blood, and has a higher rate of loss of urinary salt (Werntz, 1963). Lockwood (1961) has shown that when <u>G. duebeni</u> is transferred from 100-175% sea water to fresh water, its urine changes from isotonic to hypotonic. This ability to rapidly change the concentration of the urine and thereby regulate salt loss is an important adaptation to an environment subject to changing salinity, such as pools which may be suddenly inundated with salt or fresh water.

W.T.W. Potts (1954) has done experiments concerning the energy requirements of osmoregulation in fresh water. He lists the factors which affect this requirement as 1) the animal's permeability, 2) the surface area, 3) the concentration of the medium, and 4) the urine and blood concentrations.

Considering the last factor, one finds that most "Although many authors use the terms isotonic as synonymous with isosmotic (and similarly, hypertonic and hypotonic), the terms are not identical. Tonicity is defined in terms of the response of cells immersed in a solution. A solution is said to be isotonic with a cell (or tissue) if the cell neither swells nor shrinks when immersed in it." (Potts and Parry, 1964).

fresh-water Grustacea, like <u>G. fasciatus</u>, produce a urine hypotonic to their blood. <u>Friocheir sinensis</u>, a freshwater grapsoid crab which returns to the sea to breed, is one of the exceptions since it produces a urine isosmotic to its blood at all external dilutions (Robertson, 1957). Potts uses this crab as an example in support of the thermodynamic advantages to be derived from producing blood-hypotonic urine.

His reasoning proceeds like this: If an animal produces V 1. of urine/hr. of a concentration U moles/1., it is losing VU moles of solute/hr. To balance this loss the animal must absorb solute from the medium. The work, W, required to transfer 1 mole of solute from concentration M to concentration B = R T ln B/M cal. Then the minimum work to balance solute loss = R T V U ln B/M cal/hr. Potts assumed the following values for a 60 gm. crab: M = 0.006 mole/1., B = 0.320 mole/1., U = 0.320 mole/1., V = 1.04 × 10⁻⁴ 1./hr., metabolic energy = 14 cal./hr. He calculated that V = P A (B-M) where P is the permeability and A is the area. By this equation, $P A = 3.312 \ 10^{-4} \ 1./hr$. By R T U V ln B/M = W, W = 0.0757 cal./hr. This value is approximately 0.5% of the total metabolic energy available. But, if B = U = 1.2 moles/ 1. (a 1.2 mole solution is isotonic with 100% sea water), W = 1.69 cal./hr., and if B = 0.320 mole/l. with U = M = 0.006 mole/l., W = 0.0187 cal./hr.

Thus, as <u>Briocheir</u> goes from sea water to fresh water, B falls from 1.2 to 0.32 moles/1., and this reduced internal concentration lowers the osmotic work from 1.69 to 0.0757 cal./hr. If this crab were able to produce a urine of optimum concentration, (isotonic to the medium, U = M) the osmotic work could be further lowered to 0.0187 cal./hr. (Potts, 1954).

Potts makes the point that in fresh water, an animal can reduce its osmotic work by as much as 90% by making its urine isotonic with the medium, but even making it hypotonic to the blood (while still several times more concentrated than the medium) is extremely efficient. This is, in fact, what most fresh-water animals do; none can produce a urine isotonic with the medium. The reduction of the concentration of the urine is of maximum efficiency energy-wise in the early phases of dilution. By lowering the urine concentration to 1/10 the concentration of the blood (ie. 5% sea water), Potts calculates that 93% of the possible saving on energy has been achieved, and the urine is still 100 times more concentrated than the medium (.05% sea water).

Potts' hypothesis has been criticized because it is based on the assumption of ideal semipermeability and total solute loss via the urine. Shaw (1959) pointed out that for many fresh-water Crustacea, most of the solute loss occurs through simple diffusion across the body sur-

face. <u>Friocheir</u> loses only about 10% of its salt loss through the urine (Krogh, 1939). Furthermore, Shaw (1961b) states that most fresh-water Crustacea are not really semipermeable, and the saving in energy by the production of hyposmotic urine is not as great as Potts calculated. Lockwood (1961) noted that, although Potts' theory attributed little energy-saving advantage to hyposmotic urine in a brackish medium, <u>G. duebeni</u> uses it in 50% or less sea water.

The comments above represent part of the disagreement concerning the order in which animals assumed the fresh-water adaptive mechanisms, and even concerning which mechanisms are basic and which are refinements. According to Beadle and Cragg (1940), the ability to produce a hypotonic urine was not the first step taken by animals in adapting to a fresh-water environment. Instead, they hypothesize, the power to actively transport salts from the medium into the blood came first, while the excretory organ still produced isotonic urine. Carcinus can regulate in dilute sea water, but not in fresh water, and may be thought of as exemplifying this stage. Eriocheir is quite similar but is found more often in fresh water because it has a somewhat better salt transport system. Parry (1954) noted that this two-stage hypothesis is acceptable only for animals with a very low permeability.

Animals without this would need a tremendous power of ion absorption to offset solute loss. Ion exchange may take place at the body surface, the gut, or the excretory organ (Ramsay, 1954). Active transport of ions by surface membranes is definitely a mechanism of hyper-osmotic regulation used by fresh-water animals, yet it probably existed in marine ancestors. <u>Cancer</u>, for example, keeps a higher concentration of K than found in the normal medium, yet possesses no osmoregulatory mechanisms.

Following the ability to actively transport salts into the blood, according to the Beadle and Gragg hypothesis, came the second stage in the transition to fresh water: the ability to conserve salt by producing a hypotonic urine, and the ability to reduce the strain of regulation by tolerating a lowering of the osmotic pressure of the blood to about one-half the value found in marine animals. Since the work of active transport depends upon the concentration gradient to be overcome, the latter ability reduces regulatory work.

Shaw(1961a) disagrees with Potts about what is most basic to fresh water adaptation. As mentioned above, Potts (1954) considers lowered blood concentration and the production of a dilute urine of greatest importance. Shaw feels that these attributes are merely refinements, and instead lists as most basic: 1) a lowered permeability and 2) a reduced external concentration at which saturation of

the high-affinity uptake mechanism is reached.

Active transport of salts was studied by Koch (1954) in isolated gills of Eriocheir. He found that they were capable of absorbing NaCl from a solution of only 1/30-1/40 the concentration of the blood. Absorption stops, however, in the absence of oxygen. Koch also showed that many basic dyes are reversible inhibitors of salt absorption, possibly because of anticholinesterase activity. Shaw (1961a) studied Na balance in Eriocheir and found that the mechanism for active uptake is saturated (working at top rate) at external concentrations as dilute as 6 mM/1.; this mechanism is adequate to balance the Na lost through the body surface. The oxidative activity of crab gill mitochondria has been studied as a function of the osmotic concentration of the medium by King (1966). She found that as the osmolarity of the medium decreased, the oxygen consumption of Callinctes sapidus gill mitochondria increased by 75% (for brackish-acclimated animals) and 35% (marine crabs). When the medium was diluted from 1.6 to 0.16 Osm., the specific activity of the gill mitochondria increased by 200-300%.

Bielawski (1964) did experiments with isolated podobranch gills of the crayfish <u>Astacus leptodatylus</u> Esch. and <u>A. astacus</u> L. The gills are permeable to water and usually allow a passive water intake, but Bielawski found

that ill permeability fell when the concentration of the medium was lowered, so that regulation was facilitated. Permeability rose as the concentration of the medium was raised. Chloride and other ions are transported across the gill membrane against a gradient. The carrier for Cl is saturated at a medium concentration of 0.2%, and at meater than 0.9% sea water the carrier is inhibited. Shaw (1960) showed that in the crayfish Astacus pallipes Lereboullet, Cl and Na transport is by two different mechanisms which may act independently of each other. 1 influx takes place in the presence of an electrochemical gradient, and can occur at a high rate at lower nedium concentrations than does Na. A Cl deficiency relative to the Na concentration causes a change from C1-C1 exchange to an exchange of Cl for a useful anion, probably bicarbonate. Salt-depletion causes Na influx by activating the Na transport system. Shaw's experiment suggests that the Na system is the more basic to the maintenance of blood concentrations. After the Na level in the blood is set, the relative concentration of Cl may then activate the Cl system to bring the Cl level up to that of Na.

The site of osmoregulation, if the area of transport in the gills is in fact discrete, may have been detected by Copeland (1963). The crab <u>Callinectes</u> is able to adapt

to fresh water from sea water. Copeland found near the afferent blood supply a patch of cells which he did not find in obligatory sea water forms, and believes that these may be the cells capable of hyposmotic regulation.

It is possible for animals to take in ions not only by transport mechanisms, but also through the food they eat. Parry (1961) studied Cl regulation in the Branchipod <u>Triops cancriformis</u> Bose and found that it could maintain a blood concentration well above the concentration of the medium. In its normal fresh water environment it is able to exclude the osmotic inflow of water while meeting salt requirements by food. In the laboratory it was able to live for six days in distilled water without food. The Entomostracans <u>Branchipus</u> and <u>Chirocephalus</u> also depend upon salt uptake from food. While <u>T</u>. <u>cancriformis</u> maintains a blood concentration of 55 m.eq./l. in fresh water, by comparison, <u>Branchipus</u> has a blood concentration of 30 m.eq./l. (Krogh, 1939), and <u>Chirocephalus</u> (Panikkar, 1941) maintains 77 m.eq./l.

<u>Triops longicaudatus</u>, also a hyper-osmotic regulator, has been shown by Horne (1966) to depend more on an active uptake mechanism to maintain its necessary internal concentration of ions. Studied in pond water, <u>T. longicaudatus</u> maintained a water to hemolymph ratio of 1:435 for Na, and 1:187 for Cl. This indicates a good regulatory mechanism.

In deionized water and with a food source, this animal lost both Na and Cl from the hemolymph, yet Ca and Mg concentrations were not significantly altered, and the concentration of K rose. <u>T. longicaudatus</u>, depending more on active uptake, is more permeable to ions than is T. cancriformis.

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Brackish-water Crustacea

Brackish water is usually considered as being between 0.5% o and 30% o saline (Potts and Parry, 1964). It may be found in such places as estuaries, salt marshes, and inland seas. It is commonly believed that as crustaceans moved from the sea to fresh water, they often did so by first adapting to the transitional environment, brackish water. There is abundant evidence to support this theory. Sometimes species of the same genus are found which represent a spectrum of regulatory abilities. They may occupy niches which represent a spectrum of salinities, such as the decreasing salinity one may observe in going from the sea up a river.

The distribution of intertidal crabs in the Brisbane River was studied by Snelling (1959). The results of this study suggest that salinity is the most important factor controlling and limiting the range of estuarine crabs. Such crabs cannot extend into a given salinity unless they possess the regulatory abilities necessary to maintain their internal concentrations there.

Two populations of an estuarine isopod, Cyathura polita Stimpson, were studied by Segal and Burbank (1963) because of the salinity differences of their natural environments. One population was found in an estuary in Massachusetts, with a tidal salinity variation of from 0.5% to 17%, and an annual temperature fluctuation of from about 2°C to 23°C. The other population was found in a spring in Florida, and lived in water of salinity 1%o, with probably less than 3°C fluctuation in temperature annually. Both populations were able to hyper-osmoregulate in 50% to 3.5% sea water. When the medium concontration was increased experimentally to 75%-150% sea water, the blood concentration was approximately isosmotic with the medium. In water ranging from distilled to 3% sea water, the estuary population was able to maintain for a minimum of 48 hours the blood concentration it had maintained at higher solution concentrations. The population from the spring, however, was unable to osmoregulate at salinities less than 3% sea water. At 32°C the estuary animals maintained about the same blood concentration at varying salinities as they had at 22°C. The osmoregulatory ability of the spring animals broke down in either high or low concentrations at 32°C. Thus even

members of the same species may differ in their regulatory and adaptive powers. It is not difficult to believe that closely related animals may come to occupy separate niches due to different degrees of physiological adaptation to salinity differences.

Many estuarine animals can exist in water approaching the salinity of fresh water, indicating the ease with which a transition to a fresh-water environment might be made by some brackish-water forms. The Black Sea barnacle <u>Balanus</u> <u>improvisus</u> occurs naturally in waters of salinity of 18%. Turpaeva and Simkina (1961) found that it is able to withstand extreme dilutions: no harmful effects were observed at 5% but at 3% growth was slowed, and most died within a month in fresh water. The amphipod <u>Ampelisca spinipes</u> Boeck was found in an estuarine environment (Nagabhushanam, 1965) and in experiments could tolerate salinities down to between 3% and .h%.

It is necessary that animals maintain the osmotic pressure of their cells, whatever the salinity of the medium. Both marine and brackish animals use organic substances to maintain osmotic pressure. In fact, Lockwood (1962) states that inorganic ions contribute only about 1/3 to 1/2 of the osmotic activity in the marine Crustacea which have been studied. Lee and Mc Farland (1962) found that serum cations and Cl accounted for 95% of the osmotic pressure.

in the mantis shrimp <u>Squilla empusa</u> Say when the animal was in normal sea water. As the medium became quite diluted, these ions contributed only 67% of the osmotic pressure; the remainder was accounted for by an increase in non-protein nitrogenous compounds. When the brackishwater <u>Carcinus maenus</u> was transferred from 100 to 40% sea water, Shaw (1958) found that organic substances were contributing to the cellular osmotic pressure, since water uptake and salt loss did not account for the isotonicity of the blood and muscle fibers.

The basic mechanisms of ionic and osmotic regulation in brackish-water Crustacea are like the mechanisms used by fresh-water Crustacea, and the reader should refer to the section dealing with fresh-water Crustacea for a discussion and comparison. The mechanism of regulation in the brackish-water prawn, <u>Metapenaeus monoceros</u>, will be mentioned here because it is unlike the mechanisms discussed previously. Gnanamathu (1966) studied this prawn in a variety of medium concentrations, and found that it increased in volume in a concentrated medium and decreased in volume in a dilute medium. Fluid pressure is involved in the mechanism of active regulation of water across the gut wall. When the medium is hypertonic, water is lost and the reduced hemolymph volume produces an increase in pressure in the distended gut, causing an ultra-

filtrate of dilute sea water from the gut into the hemolymph. The gut may alter the body volume, hence pressure, by the influx or efflux of water. In a hypotonic medium, the entry of water may put pressure on the gut and check further entry of water, besides facilitating filtration of hypotonic water from the hemolymph into the gut.

Marine Crustacea

The blood of most marine Crustacea, except prawns. isopods, and grapsoid crabs, is isosmotic to sea water (Lockwood, 1962), but almost nothing is known of osmoregulation in marine ostracods, copepods, cirripeds, mysids, cumaceans, or euphasiids (Robertson, 1960). Those animals which are in osmotic equilibrium still must use ionic regulation because sea water does not have all ions in the same proportions as they are needed in the body fluids of marine animals. This regulation takes place by basically two mechanisms: 1) selective excretion of ions by the renal organs, and 2) controlled uptake of ions with water from the medium. This uptake may be either by absorption against a gradient, or perhaps by swallowing sea water and excreting the excess salt across the gills, as is done by teleosts. Gross (1955) showed that P. crassipes is able to absorb water against a gradient.

In general, marine Crustacea produce a urine nearly

isosmotic with the blood. The antennal glands, then, are not concerned with osmoregulation but rather with controlling the ionic composition of the blood. Regulation usually involves maintaining the internal concentration of Na, K, and Ga above the concentration found in sea water, keeping the Mg and SO₄ concentrations lower than in sea water, and maintaining the Cl concentration close to equilibrium (Robertson, 1960). When the Mg and SO₄ concentrations are lower than found in sea water, the ionic concentration and cation-anion balance in the body fluids must be corrected. This is usually accomplished by raising the Na concentration and the Cl concentration, respectively.

A study by Dehnel and Carefoot (1965) dealt with blood and urine concentrations determined both in winter and summer for <u>Hemigrapsus mudus</u> and <u>H. oregonensis</u>. They detected no significant difference between winter and summer values for the blood ion concentration for Na over a range of 6% to 175% sea water; no difference for the K concentration over most ranges, except for being higher in the summer at a medium concentration of 100% to 150% sea water; a higher value for Ca in the winter than in the summer; and no significant difference for Mg. The summer urine Na concentration was approximately the summer blood Na concentration, indicating that an extra-renal mechanism is responsible for concentrating Na in the body. The summer urine K concentration also was about equal to the blood K concentration, thus the antennary glands are shown to be ineffective for the regulation of K. In fact, regulation of ion concentration was shown to be extra-renal for all ions but Mg. The antennal.gland is used to maintain a hypotonic Mg concentration. This ability.was impaired by increased temperatures.

Parry (1953) found that the isopod <u>Ligia oceanica</u> maintains an efficient ion regulatory system which keeps Na, K. Ca, and Cl concentrations in the blood hypertonic to sea water. The salts are probably derived from food or absorbed directly from the medium at some body surface such as the pleopods. The osmotic pressure of the blood can be regulated in salinities of from 20% to 100% sea water, but the blood concentration conforms to the medium concentration at values less than or greater than this, indicating a failure in the regulatory mechanism at these concentrations.

Research on ionic regulation in the palaemonid prawn <u>P. serratus</u> led Parry (1954) to hypothesize a regulatory mechanism involving an ultra-filtrate system. This prawn is normally hypotonic to sea water, with the blood concentration of Na, K, and Cl approximately 70-80% of the concentrations found in sea water. The Mg and SO₄ concentrations are lower in the blood than in sea water, and higher

*See note on page 11. This writer used tonicity if it was used by the author of the work cited.

In the urine than in the blood. The urine is isotonic to the blood, and because the Na, K, Cl, and Ca concentrations are about equal in the two solutions, Parry suggested that the urine is an ultrafiltrate of the blood. Because Ng and SO_{μ} concentrations were higher than in the blood, she suggested that these ions were perhaps actively excreted after formation of the ultrafiltrate. The antennal cland of the palaemonid prawns does not have a site such as the tubule of fresh-water crayfishes where saltabsorption and osmoregulation take place. Parry states the the end-sac of the antennal gland may be the site of the ultrafiltrate production, and active excretion of Mg, SO_{μ}, and perhaps MR_3 may-occur around the nephridial canal. Beabsorption of ions to balance charges might take place here also.

Riegal (1959) measured the changes in the total osnotic concentration of body fluids in two spheromid isopods, <u>Gnorimi spheeroma oregonensis</u> Dana and <u>Spheer. pendoton</u> Richardson, after exposure for 3 to 48 hours in varying salinities. In less than 50% sea water the animals were hypertonic; from 75% to 125% sea water, they were usually hypotonic. Because weight changes were not observed when the internal concentration changed, the change took place by salt movement. If the temperature was allowed to drop from 16°C to 5°C there was a marked increase in weight which suggests that under optimum temeratures the total water content is actively maintained, and that low temperatures must destroy the effectiveness of that mechanism. The change in concentration observed at 5°C could not be explained only by the increase in water volume, therefore salt movement was still taking place.

Terrestrial Crustacea

In a terrestrial habitat an animal is continually losing water from its body by evaporation and in the urine. Loss of water from body fluids means that the internal concentration of ions will tend to rise, and may in this way exceed in time the optimum concentration required for proper tissue functioning. Continued loss of water may result in death. To ensure that the body fluids do not become too concentrated, land forms have adopted ways to reduce water loss. 'Means by which this may be accomplished include the following:

1) Semi-terrestrial and terrestrial crustaceans have an integument that is less permeable than is the integument of marine forms (Edney, 1960). This condition prevents great losses from the general gody surface.

 Land forms tend to reduce the surface area of bissues used for gas exchange, eg. the gills (Nicol, 1960), since so much evaporation takes place from the moist,

exposed tissues.

 Terrestrial animals in general produce a rather small volume of urine.

Because these measures cannot eliminate all water loss, the animal may also seek to maintain its internal concentration by taking in water to dilute the ions which do become concentrated. To take up water, terrestrial orustaceans may eat moist food, drink from pools, enter water, or absorb water from sand. <u>Gecarcinus lateralis</u> is a brachyuran crab which lives a terrestrial life and is able to live indefinitely simply by exposure to sand dampened with fresh water (Gross, 1963). It is able to use the fresh water absorbed from the sand to dilute its body fluids, for it dies if sea water is the only source from which the sand is dampened. Flemister (1958) found that <u>Gecarcinus</u>, well removed from the marine life and living above the intertidal zones, was intolerant to either sea water or sand dampened with sea water.

Cardisoma carnifler, another brachyuran, can take up water against a gradient from sand dampened with either sea water or fresh water. The advantage to be gained in adaptation to terrestrialness may be questionable in this case, however, since the animal produces a urine hyposmotic to the body fluids after such rehydration, thereby losing water (Gross et. al., 1966). It is believed that terres-

trial forms evolved from marine forms, and <u>Cardisoma</u> and <u>Gecarcinus</u> represent different degrees of evolution away from a marine life. <u>Gardisoma</u> lives in the intertidal and above, and will enter sea water and tolerate high salinities. <u>Gecarcinus</u> lives away from the water, and, as mentioned above, is intolerant to sea water. <u>Gecarcinus</u> may even drown in water, which <u>Cardisoma</u> will not do (Gross et. al. 1966).

Behavioral means of controlling water losses are well-documented. Cardisoma guanhumi Latreille is very independent of the water, but may enter sea water or fresh water to take in water (Gifford, 1962). The anomuran land crab Coenobita cavipes was shown by Gross et. al. (1966) to be able to control its water balance by behavioral means. Pachygrapsus crassipes, which may be considered to be a semi-terrestrial crab, has been shown to prefer 100% sea water (Gross, 1957) to other salinities. Because it can detect and avoid abnormal salinities, it can sometimes prevent the osmoregulatory problems associated with dilute or concentrated pools. This mechanism may restrict its habitat to the intertidal and subtidal zones rather than allow more freedom in range. Coenobita perlatus will spend 90% of its time out of the water if allowed a choice (Gross and Holland, 1960). It prefers to visit sea water five times more often than fresh water, when it

coes choose to enter water, possibly because its internal concentration is not so high as to necessitate dilution by fresh water. This assumption is supported by the fact that, if its internal concentration is forced away from the normal value, it seems to be able to select water having a salinity which might best restore its internal concentration.

The role of food in affecting the internal concentration by contributing water was mentioned above. Food may also add ions. Gross and Holland (1960) studied the terrestrial hermit crab Coenobita perlatus under a variety of experimental conditions, including fresh water; fresh water and food; fresh water and sea water; sea water and food; fresh water, sea water, and food; and 150% sea water and food. After 12 days in the experimental environments, the crabs were analyzed for blood and urine concentrations. Then given fresh water and food, the blood Na, K, and Mg remained fairly constant, regardless of whether or not sea water was accessible. Given both sea water and fresh water, en animal was able to maintain its internal concentration without the aid of food. Given only fresh water, however, the blood was less concentrated than when both food and fresh water were available.

Because of the difficulties in precisely controlling the internal concentrations, terrestrial forms may find

it advantageous to be able to tolerate wide fluctuations in the body fluid concentration. <u>Cosmobile perlatus</u> can tolerate a Na concentration range of 83% to 220% of the normal value (Gross and Holland, 1960).

Terrestrial crustaceans are usually good hyposmotic regulators. Gross (1964) studied seven species of orabs, from a range of habitats, to determine their relative abilities to regulate. The crabs that he studied were the equatic <u>H</u>. <u>oregonensis</u> and <u>Cancer antennarius</u>; the semi-terrestrial <u>P</u>. <u>orassipes</u>, <u>Grapaus grapsus</u>, <u>Uca cremulata</u>, and <u>Coypode ceratopthalma</u>; and the terrestrial <u>Gecarcinus lateralis</u>. He found a definite correlation between the degree of terrestrialness of these crabs and their relative abilities as hyposmotic regulators.

The role of the antennal gland in ionic and esmotic regulation has been the subject of much research. Although both <u>Cardisoma carnifex</u> and <u>Sesarmi meinerti</u> were shown by Gross et al. (1966) to be strong hyper-osmotic and hyposmotic regulators, the antennal gland does not seem to be osmoregulatory in function, since the urine was isosmotic with the blood. It can help in ion regulation, however, by concentrating Mg in the urine independent of the Mg influx from the medium, and probably at the expense of Ma. The ability to regulate Mg is not necessarily correlated with a low concentration of Mg in the blood, for Uca crenulata was the best of the seven crabs studied by Gross (196)) at concentrating Mg in the urine, yet it has a blood concentration of Mg as high as does Cancer, with a poor ability. Gecarcinus was found by Gross (1954) to be an exception to the other terrestrial crabs in his study, because it has only a poor ability to regulate Mg. He hypothesized that Gecarcinus may have followed a different evolutionary path of physiological adaptation to terrestrialness. If so, Coenobita perlatus has probably done so also. In experiments with Coenobita perlatus, Gross and Folland (1960) found that when the salinity of the external medium was increased, the concentration of the blood Na and Mg increased, K increased somewhat, and Ca did not increase at all. They concluded that the antennary glands are poor ion regulators. Coenobita may depend more on its high tolerance for variation in its internal concentrations than on ion regulation to live a terrestrial life. Gross (1959) studied the ion-regulating powers of the antennal gland of P. crassipes. It was ineffective in the regulation of Na, K, and Ca, but much more effective in the regulation of Mg. Gross et al. (1966) presented evidence of conservation of Ca by the antennal glands of C. carnifex, and S. meinerti by showing that the ratio of the urine Ca to the blood Ca was less than 1. In general, the antennal fland of terrestrial crustaceans is ineffective for osmoregulation, but can function in limited ion regulation, most conspicuously for Mg.

Summary and Conclusions

Canotic and ionic regulation in Grustacea has enabled various members of the Grustacea to leave a marine envinonment and to adapt successfully to brackish, frosh-water, and terrestrial habitats. It has been shown that many animals are limited in range basically due to a limited ability to adapt to different salinities.

Fresh-water Grustacea generally have a low permeability to water and salts. This condition lessens the dilution of their body fluids by means of outward diffusion of salts and water intake by osmosis. Ion absorption takes place primarily across the gills by active transport. The energy cost of this transport is proportional to the concentration gradient existing between the internal body fluids and the medium. By acquiring a toleration for less concentrated internal fluids than their marine ancestors, fresh-water forms reduce the energy cost of active transport. It is to the advantage of fresh-water animals to have ion carriers which saturate at very low external concentrations. Freshwater animals also benefit from producing a dilute urine.

Brackish-water crustaceans are more permeable than their fresh-water relatives. The concentration of their body fluids is in general greater than the concentration of the medium. Existence in a brackish medium may be possible either because the animal is able to regulate its internal concentration of ions or because it can tolerate large changes in the concentration of its body fluids.

Marine animals tend to be isosmotic with sea water. There is thus little need of osmoregulation, but the need for ionic regulation still exists. There is active ionic uptake by the gills, and the antennal glands have a limited role in ion regulation. Some crustaceans are able to actively maintain a hyposmoticity in sea water. The means by which this is accomplished is deserving of more research.

Terrestrial crustaceans must cope with a high rate of water loss through evaporation and via the urine. It is to their advantage to be quite impermeable. Water loss is partially balanced by drinking, eating moist food, and behavioral modes such as immersion in pools. Many terrestrial forms are able to regulate hyposmotically, while others have adapted by toleration of wide fluctuations in their internal concentrations.

The physiological and behavioral means to regulate osmotically and ionically have been the subject of a great deal of research. Although a large body of information has accumulated dealing with decapods, there is a paucity

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of information available on many of the other crustacean roups. Research needs to be carried out on these forms in order to obtain a more clear understanding of the possible variety of regulatory mechanisms. Active transport is fundamental in any consideration of ionic or camotic regulation. The carriers involved in active transport are not yet positively identified, and more research on the mistology and biochemistry of exchange membranes is needed.

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